

Impact of Treatment with Imatinib on Renal Histology in Albino Rats

*Luma Ibrahim Khalel Al-Allaf, Hafidh Al-Ashoo **

Abstract

Background: Targeted small molecule drugs have revolutionized treatment of chronic myelogenous leukemia (CML) over the last decade. However, their use has been found to be associated with serious toxic effects on a number of vital organs including the kidneys.

Objectives: This study aims to determine the histological changes of the kidney of rats after administration of a dose of 75mg/kg/once/day of Imatinib mesylate for one month in comparison to control ones.

Study setting and design: This experimental study was conducted on 16 male Albino rats purchased from Animal Houses of Veterinary College, University of Mosul, Mosul, Northern Iraq.

Methods: In this study a group of eight rats (40-45 days) were administered orally daily dose of 75mg/Kg/30 days of imatinib mesylate (Glivec[®], Novartis). Another group of 8 rats (40-45 days) were administered distilled water (D.W). Kidneys tissues from each rat were obtained. The tissues were embedded in paraffin and stained with hematoxylin-eosin, periodic acid schiff + Hematoxylin stain, Toluidin blue, and Masson's Trichrome.

Results: Rats treated with 75mg/kg /once/day of imatinib for 30 days showed different histological changes in glomeruli and some parts of the urinary tubules in comparison with controls. The most evident features are increase in Bowman's space, presence of lobulated or segmented glomeruli, shrunken glomeruli, dilated tubules with sloughed epithelium, and cloudy degeneration. Congested glomerular capillaries are also noticed sometimes in these sections with decreased Bowman's space. Dramatic renal injury in these rats was represented with tubular cell swelling, loss of brush border of proximal convoluted tubules as well as presence of cloudy degeneration of tubules. In addition, there was a focal accumulation of inflammatory cells of early inflammation that infiltrate between the tubules at the cortical and corticomedullary portion and early fibrosis is noticed. Sections obtained from rats treated with imatinib exhibit dilatation and hyperemia in the intertubular cortical or juxtamedullary blood vessels with appearance of structureless eosinophilic area of necrosis. In addition, dilated tubules with accumulation of eosinophilic homogenous material in tubular lumen were noticed. The interstitial tissue showed area of hypercellularity, interstitial oedema and infiltration of mononuclear inflammatory cells (lymphocytes) which tend to be concentrated around the tubules in the cortical and medullary zones.

Conclusion: Imatinib has adverse effects on the renal histology and results in alterations in the renal cortex glomerular cells or tubular, which could play an important role in renal dysfunction. A clinical collaboration between oncologists and nephrologists could be useful with the objective to optimize the management of tyrosine kinase inhibitors.

Keywords: Imatinib mesylate, chronic myelogenous leukemia, nephrotoxicity, albino rats.

(J Med J 2020; Vol. 54(3):99-114)

Received

September 28, 2018

Accepted

March 3, 2020

*Assistant Professor, Department of Anatomy, Mosul College of Medicine, University of Mosul, Mosul, Iraq.

Correspondence to:

Luma Ibrahim Khalel Al-Allaf

Department of Anatomy, College of Medicine, University of Mosul, Mosul, Iraq, E-mail: lumaallaf1971@yahoo.com

Introduction

Targeted small molecule drugs have revolutionized treatment of CML over the last decade. However, their use has been found to be associated with serious toxic effects on a number of vital organs including the kidneys^(1,2).

Chronic myelogenous leukemia (CML) is a myelo- proliferative disorder characterized by the presence of translocation t(9;22)(q34;q11) which generates the Philadelphia (Ph⁺) chromosome and the associated fusion gene (Abelson murine leukemia-break point cluster region)(BCR-ABL)⁽³⁾, which has deregulated tyrosine kinase activity and leads to increased cellular proliferation, resistance to apoptosis and genetic instability and it is at the center of CML pathogenesis⁽⁴⁾. CML, once considered a fatal disease, is now essentially a chronic disorder, and most patients can enjoy long-term survival. This history of success has been the result of development of TKIs, compounds which suppress the abnormal tyrosine kinase (TK) activity of the BCR-ABL1 protein^(5,6).

Imatinib (Gleevec® or Glivec® Novartis, NJ), is a selective, rationally designed, c-KIT and Bcr-Abl tyrosine kinase inhibitor, approved for the treatment of chronic myelogenous leukemia (CML)⁽⁷⁾, gastrointestinal stromal tumors (GIST)^(8,9), and unresectable GIST⁽¹⁰⁾.

In addition, imatinib inhibits the platelet derived growth factor (PDGF) receptor,⁽¹¹⁾ which may allow further therapeutic applications including dermatofibrosarcoma protuberans⁽¹²⁾, glioblastoma⁽¹³⁾, and non-cancer related pathologies like rheumatoid arthritis⁽¹⁴⁾, and atherosclerosis⁽¹⁵⁾.

Imatinib undergoes P450 mediated metabolism mainly via CYP3A4 and CYP3A5 which play a minor role⁽¹⁶⁾. Imatinib and metabolites are excreted in the bile and only around 5-12% is excreted unchanged in urine

^(17,18). The main adverse effects include severe neutropenia and thrombocytopenia, oedema, fluid retention, nausea, mild diarrhoea, skin rashes, arthralgia, myalgia, bone pain, acute renal failure and hepatotoxicity^(19,20,21,22,23).

Renal dysfunction is often unrecognized by the treating physicians who usually base their diagnosis on serum creatinine levels which is not a sensitive estimator of renal function and may give the physician the wrong impression that the renal function is still normal⁽²⁴⁾. Therefore, the use of formulas to estimate the glomerular filtration rate (GFR) or other methods that measure GFR is crucial and should be carried out routinely⁽²⁵⁾. There is little data on the influence of imatinib especially on the kidneys' histology.

The rat is one of the most widely used research animal particularly the urinary anatomy, histology, and physiology⁽²⁶⁾. The rat is also useful in assessment of toxicological insult to the urinary system. They are initially used for experimental purposes since the half of the nineteenth century. Several strains have been developed for studying several diseases^(27,28,29).

This study **aims** to evaluate the histopathological changes that occur in kidneys of rats after treatment with low dose of imatinib mesylate (75mg/kg/once/day) for one month duration in comparison to the control ones.

MATERIALS AND METHODS

This experimental study was conducted on male Albino rats purchased from Animal Houses of Veterinary College, university of Mosul, Mosul, Northern Iraq. Throughout the investigations the rats were housed under controlled normal environmental laboratory conditions and animal facility. They were local bred and put individually in Animal House

plastic cages^(30,31) and provided with free access of water *ad libitum* and pelleted food⁽³²⁾. The experiments were performed during the light portion⁽³³⁾.

Experimental design and procedures

Mean bodyweight of all rats was 70-110 gm. The first experiment includes 40- 45 days aged rats which were administered daily dose of 75mg/Kg of imatinib mesylate (Glivec® Novartis) purchased from IBN-SENA Teaching Hospital or bought from some private pharmacies and were dissolved in D.W and were administered orally by gavage with needle (24 G) for 30 days (n=8) with age matched control who administered D.W following the same protocol applied to imatinib group (n=8).

Imatinib doses selected were intended to be in the range of those used in clinical treatment regimens⁽³⁴⁾. (400-800 mg/d or 340-590 mg/m² based on a weight of 70 kg) dose surface area adjusted to body-weight, $f \times \text{mg/kg} = \text{mg/m}^2$, where f is a constant equal to 6.0 in rats⁽³⁵⁾. Each animal was observed for overt signs of toxicity. The animals were firmly restrained (the animal was grasped by the loose skin of the neck and back) to immobilize the head and maintained in an upright (vertical) position. The needle (24 G) was passed through the side of the mouth, followed the roof of the mouth, and advanced into the esophagus toward the stomach. After the needle was passed to the correct length, imatinib was injected⁽³⁶⁾.

Study termination procedures: Animals in each experiment were euthanized with ether^(30,37). 24 h after the final dose was given.

Tissue and organs collection: Kidneys of rats from each experimental group were obtained using longitudinal thoracoabdominal incision. The Kidneys were excised and examined macroscopically.

Preparation of histological sections:

Kidneys were fixed in 10% Neutral buffered formalin⁽³⁸⁾. The frozen embedded wax blocks (Merck, Germany) were sectioned at 3-5 μ thickness using Reichert-Jung microtom (Austria)^(39,40), and slides were stained with Harris hematoxylin-eosin (Scarlau, Spain) for general renal structure, periodic acid schiff stain+Harris hematoxylin (PAS+H) to demonstrate the glycogen deposition in these sections, Toluidin blue stain was used as routine stain for renal sections, while Masson's Trichrome were used for detection of mitochondrial contents in sections. The evaluation was blinded to treatment and any data by an expert histopathologist (Prof.Dr. Al- Nuami's WMT).

Histopathological analysis

Renal changes were graded as mild, moderate, or severe. Scores +, ++, and +++ are mild, moderate, and severe levels, revealing less than 25, 50, and 75% histopathological alterations of total fields examined, respectively⁽²³⁾.

Photography:

All sections were visualized in Bright field Olympus microscope (Japan). Photomicrographs of representative changes were taken using digital camera (Optika, Italy, HD 1080, resolution 8.0 Mega pixels) attached using plan apochromatic objectives. The magnifications of photomicrograph were indicated with the legends for the photograph.

RESULTS

At necropsy, no obvious gross tissue abnormalities were noted in the kidneys of any animal.

The renal section of control rat (with intake of D.W alone) showed normal architecture of renal glomeruli with intact Bowman's capsule. Brush bordered cuboidal epithelium lining the

proximal convoluted tubules. Simple cuboidal epithelium lining the distal convoluted tubules and normal medullary renal tubules (**Figures 1a&b**).

Rats treated with 75mg/kg /once/day of imatinib for 30 days showed different histological changes in glomeruli and some parts of the urinary tubules in comparison with controls. The most evident features are increase in Bowman's space, presence of lobulated or segmented glomeruli, shrinkage of glomeruli, dilated tubules with sloughed epithelium, and cloudy degeneration (**Figure 2**). Congested glomerular capillaries are also noticed sometimes in these sections with decreased Bowman's space (**Figure 2**). Dramatic renal injury in these rats was represented with tubular cell swelling, loss of brush border of proximal convoluted tubules as well as presence of cloudy degeneration of tubules.

In addition, there was a focal accumulation of inflammatory cells of early inflammation that infiltrate between the tubules at the cortical and corticomedullary portion (**Figure 3**). While early fibrosis is noticed in **Figure 4**. Sections obtained from rats treated with imatinib exhibit dilatation and hyperemia in the intertubular cortical or juxtamedullary blood vessels (**Figures 5,6**), with appearance of structureless eosinophilic area of necrosis as shown in **Figure 5**. In addition, dilated tubules with accumulation of eosinophilic homogenous material in tubular lumen were noticed in **Figure 7**.

The microscopic observations in **Figure 7** showed hyperemic vessels, congestion and dilatation of vessel, between the degenerated renal tubules at the corticomedullary portion. Dilated tubules with flattened epithelium, presence of sloughed cells in their lumina and extravasated RBC were revealed in **Figure(8)**.

While **Figure (9)** showed sections with thickening of blood vessel wall.

On the other hand, the interstitial tissue showed area of hypercellularity, interstitial oedema and infiltration of mononuclear inflammatory cells (lymphocytes) which tend to be concentrated around the tubules in the cortical and medullary zones (**Figure 10**).

The effect of imatinib on renal histology in study groups is shown in **Table 1**.

Sections which were obtained from the rats of the control group showed presence of a considerable amount of carbohydrates in the cytoplasm of kidney cells using PAS-technique, which gave a red or magenta colour (**Figures 11a and 11b**). The nuclei, however, appeared entirely PAS-negative staining, indicating absolute lack of carbohydrates.

Treating rats with imatinib, caused a decrease of total carbohydrates in the kidney cells (**Figure 12**).

Using Masson's Trichrome stain **Figure 13a&b** showed the normal reaction of the renal cortex and medulla.

Sections obtained from rats treated with imatinib revealed increase in the reaction to Masson's Trichrome stain with decrease in the mitochondrial contents compared to that of control rats (**Figure 14**).

Effect of imatinib on the reaction of the renal sections of different groups to PAS is shown in **Table 2**.

DISCUSSION

Tyrosine kinase inhibitors are not entirely BCR-ABL1-specific, and this lack of specificity could account for the off-target effects of these drugs^(25,41). It has been reported that these adverse events are off-target effects that are detrimental to the patient⁽⁴²⁾.

The present study indicated that

administration with 75mg/kg of imatinib induced various histopathological alterations in the renal sections of rats. These findings are consistent with those of others^(43,44), where they classified the drug-induced renal disease into 3 main areas- glomerular injury, vascular injury and tubule-interstitial changes. Observations of previous studies revealed that lesions in these areas are frequently seen together as shown in this study^(43,45).

This work revealed that imatinib has an influence on renal histology in these different areas with approximately similar degrees. These observations were similar to that of others⁽⁴¹⁾, who reported that the renal toxicity of targeted therapies are most often due to structural damages of the nephron. In addition, several case reports showed that coincidentally, with the start of treatment with imatinib, the patient develop acute renal failure^(2,46,47), and acute tubular necrosis as being observed on histopathology⁽²⁾.

However, Nassar *et al.*, showed that there are very minor renal changes were observed, which may be anticipated since they used a single oral dose of 100 mg/kg of imatinib⁽³⁹⁾.

Light microscopic observations in the current study showed that repeated administration of imatinib at 75mg/kg induced several lesions as the appearance of glomerular swelling, periglomerular fibrosis, peritubular fibrosis, accumulation of mononuclear inflammatory cells in renal cortex and medulla, interstitial oedema and necrosis. Marcolina *et al.* revealed that the long-term treatment of imatinib may cause a clinically relevant decrease in the estimated glomerular filtration rate- GFR⁽²⁵⁾. However that is consistent with those of others, who treated a group of mice with a single dose of 100mg of imatinib⁽³⁹⁾. Padmini and Kumar reported that these findings

reflect the severity of renal injury⁽⁴⁸⁾.

In addition, Alwin and Arthur reported that sloughed epithelial cells, cell debris and casts formation are a frequent finding of drug-induced renal injury⁽⁴³⁾.

The exact mechanism of chemotherapy-induced nephrotoxicity is not yet completely understood^(21,49,23,50). Some authors suggested that this renal adverse effect may be caused by two possible mechanisms: the first is tumor lysis syndrome, with precipitation and deposition of uric acid in the renal tubules, and the second is the toxic tubular damage⁽²⁵⁾. Tubular cells are susceptible to the toxic effects of drugs, as they have a role in concentrating and reabsorbing the glomerular filtrate, what exposes them to high levels of circulating toxins⁽⁵¹⁾. In the case of imatinib, the toxic effect may be related to platelet-derived growth factor receptor (PDGFR) inhibition^(52,22).

Platelet-derived growth factor b-chain (PDGF-b) expression has been reported in proximal tubules and mesangial and interstitial cells⁽²⁵⁾. It has been shown in animal models that PDGF-b/PDGFR axis plays an important role in renal tubular cell regeneration after acute tubular necrosis⁽⁵²⁾. So, by inhibiting PDGFR, imatinib may interfere in tubular repair mechanisms.

The cytokines and tyrosine kinase receptors inhibited by imatinib and other tyrosine kinase inhibitors are important regulators of the two main mechanism of tissue injury repair, regeneration, and fibrosis⁽⁵³⁾.

On the other hand, Marcolin *et al.*, revealed that the introduction of imatinib therapy in nonclinical trial CML patients is associated with decrease in estimated GFR and potentially irreversible acute renal injury⁽²⁵⁾.

Recently, renal associated adverse effects (RAEs) associated with sunitinib (a second

generation of TKIs) were reported in three studies. Two of them are clinical ^(49,50), while another is an experimental by Lim *et al.*, 2010, who revealed presence of tubular necrosis in renal sections after sunitinib administration to a group of mice. Baek *et al.*, 2013 reported severe cases of nephrotic syndrome, thrombotic microangiopathy, and acute kidney injury after administration of sunitinib ^(50,54).

This renal induced toxicity may be part of a multi organ damage mediated mainly through free radical formation eventually leading to membrane lipid peroxidation oxidative stress ^(37, 55). Induction of apoptosis and modulation of nitric oxide (NO) are other mechanisms that may be involved in toxic adverse effects associated with such therapy ⁽²³⁾.

The toxic effects to other organs as the heart and the liver may modulate blood supply to the kidney and alter xenobiotic detoxification processes, respectively, thus indirectly contributing to drug-induced nephropathy ^(25,56).

The results from a study of Hu *et al.*, 2011 ⁽⁵⁷⁾, strongly suggest that imatinib induces cardiac dysfunction through disruption of autophagy and induction of ER stress, independent of c-Abl inhibition, while Saad *et al.*, reported that the effect of imatinib on rat's livers may be attributed to the increase in NO production ⁽³⁷⁾. Imatinib induced cardiotoxicity might be attributed to imatinib- induced PDGF receptor and c-Abl blockade ⁽³⁷⁾. Recently, Hassan and Yousif reported that imatinib induce oxidative stress and release of cytochrom C and activation of caspases and leading to apoptosis in cardiac tissue of male rabbits ⁽⁵⁶⁾.

Research on the pathogenic mechanisms of imatinib-induced hepatotoxicity or renal toxicity suggests that toxicity may be related to the P450 mediated metabolic pathway or idiosyncratic reactions in susceptible

individuals ⁽⁵⁾.

The mechanism of glomerular and tubular injury such as swelling of their lining epithelial cells starts as a decrease in O₂ levels which causes a drop in aerobic respiration. Renal cells consume oxygen at a high rate and are highly dependent on aerobic metabolism for ATP production ⁽²¹⁾, the cells must rely more on glycolysis. Glycolysis leads to lactic acid builds up which causes the intracellular pH to drop. Persistent ischemia can lead to mitochondrial and lysosomal damage, and membrane damage ⁽⁵⁸⁾.

Another prominent histopathological changes in imatinib exposed kidney is thickening and interruption of glomerular basement membrane, which may play a role in renal dysfunction and reflected the decreasing in the GFR ^(44,48).

Light microscopic investigations of the current study showed a prominent dilatation of the renal vessels, authors considered that finding as one of the structural changes of kidneys ⁽⁵⁹⁾. It has been suggested that excessive production of NO causes vasodilatation and hypotension leading to organ hypoperfusion, edema and organ dysfunction ⁽³¹⁾.

In the present investigation, many renal tubules of the rat kidneys showed marked degenerative lesions under the effect of imatinib. This is justifiable since the renal tubules are particularly sensitive to toxic influences, in part because they have high oxygen consumption and vulnerable enzyme systems ⁽⁵⁸⁾. Also the tubules come in contact with toxic chemicals during their excretion and elimination by the kidneys ⁽²⁵⁾. Such degenerative changes were markedly pronounced in the proximal convoluted tubules, these findings are similar to that of Padmini and Kumar ⁽⁴⁸⁾. In addition, disintegration of

brush border membrane was shown in the present study, which may be responsible for the observed renal dysfunction⁽⁴⁴⁾. Damage to the brush border and leakage of alkaline phosphates (ALP) and gammaglutamyl transferase (GGT) enzymes, which are associated with the brush border of the renal tubules, as a result of toxin binding to the brush border and considered as an early marker of toxic tubular insult.

The results showed that treated rats with imatinib caused a depletion of carbohydrates in the cytoplasm of renal tubules. This result was in correspondence with other studies reported by others⁽⁶⁰⁾. Finally, the findings of an increase of connective tissue elements of the kidney are similar to that of others⁽⁵⁹⁾.

In conclusion: Imatinib has adverse effects on the renal histology and results in alterations in the renal cortex glomerular cells or tubular, which could play an important role in renal dysfunction. A clinical collaboration between

oncologists and nephrologists could be useful with the objective to optimize the management of tyrosine kinase inhibitors. Attention must be paid to concomitant administration of other potentially nephrotoxic agents, to avoid additive nephrotoxicity in these patients.

ACKNOWLEDGMENT

The authors would like to thank the staff of Ibn-Sena Teaching Hospital for their help in facilitating this work. Thanks are due to assistant professor Dr. banan Al-bagoo, and to Professor Dr. Wahda Al-nuaimi, department of Pathology, College of Medicine, University of Mosul and to assistant professor Dr. Mohammad Shindala, department of pharmacology, Veterinary College, University of Mosul for their great help. We are so grateful to Miss Lumyaa Zaghloul, Department of Anatomy, College of Medicine, for her excellent comments.

Table 1. The effect of imatinib on renal histology in both groups.

Group		Control N(%) Mean \pm SD N=8	Imatinib N(%) Mean \pm SD N=8	P-Value
Lesion				
Disruption of normal architecture		0(0.0%)	2(25.0%)	P<0.05
Glomerular Lesions	Increased Bowman's space	1(12.5%)	5(62.5%)	P<0.05
	Glomerular segmentation/lobulation	1(12.5%)	4(50.0%)	P<0.05
	Loss of epithelium of Bowman's capsule	0(0.0%)	2(25.0%)	P<0.05
	Glomerular hyalinization	0(0.0%)	2(25.0%)	P<0.05
	Hypertrophy of glomerular cells	0(0.0%)	1(12.5%)	P<0.05
	Glomerular Edema	0(0.0%)	2(25.0%)	P<0.05
	Glomerular congestion /disruption of capillaries	0(0.0%)	2(25.0%)	P<0.05
	Glomerular shrinkage	0(0.0%)	2(25.0%)	P<0.05
	Apoptosis	0(0.0%)	1(12.5%)	P<0.05
Tubular Lesions	Tubular dilation	0(0.0%)	3(37.0%)	P<0.05

Group		Control N(%) Mean \pm SD N=8	Imatinib N(%) Mean \pm SD N=8	P-Value
Lesion				
	Tubular edema	0(0.0%)	2(25.0%)	P<0.05
	Tubular necrosis	0(0.0%)	2(25.0%)	P<0.05
	Tubular degeneration	0(0.0%)	3(37.0%)	P<0.05
	Desquamated Tubular cells	0(0.0%)	2(25.0%)	P<0.05
	Loss of brush border	0(0.0%)	1(12.5%)	P<0.05
	Peritubular infiltration with inflammatory cells	0(0.0%)	2(25.0%)	P<0.05
	Apoptosis	0(0.0%)	1(12.5%)	P<0.05
	Detached tubules from basement membrane	1(12.5%)	1(12.5%)	P<0.05
	Tubular cast	1(12.5%)	3(37.0%)	P<0.05
	Tubular cells hypertrophy	0(0.0%)	3(37.0%)	P<0.05
Interstitial Lesions	Dilated blood vessels	1(12.5%)	4(50.0%)	P<0.05
	Hemorrhage	0(0.0%)	2(25.0%)	P<0.05
	Interstitial edema	1(12.5%)	3(37.0%)	P<0.05
	Interstitial Infiltration with inflammatory cells	0(0.0%)	3(37.0%)	P<0.05

*P-Value is considered as significant if it is <0.05.

Table 2. Effect of imatinib on the reaction of the renal sections of different groups to PAS.

Groups	Control N=8	Imatinib N=8
Parameter		
Glomerular cells	+	+/-
Glomerular basement membrane	+	+/++
Tubular cells	+	+/-
Tubular basement membrane	+	+/-
Interstitial tissue	+	+/-

* ++, moderate; +, mild; -, negative.

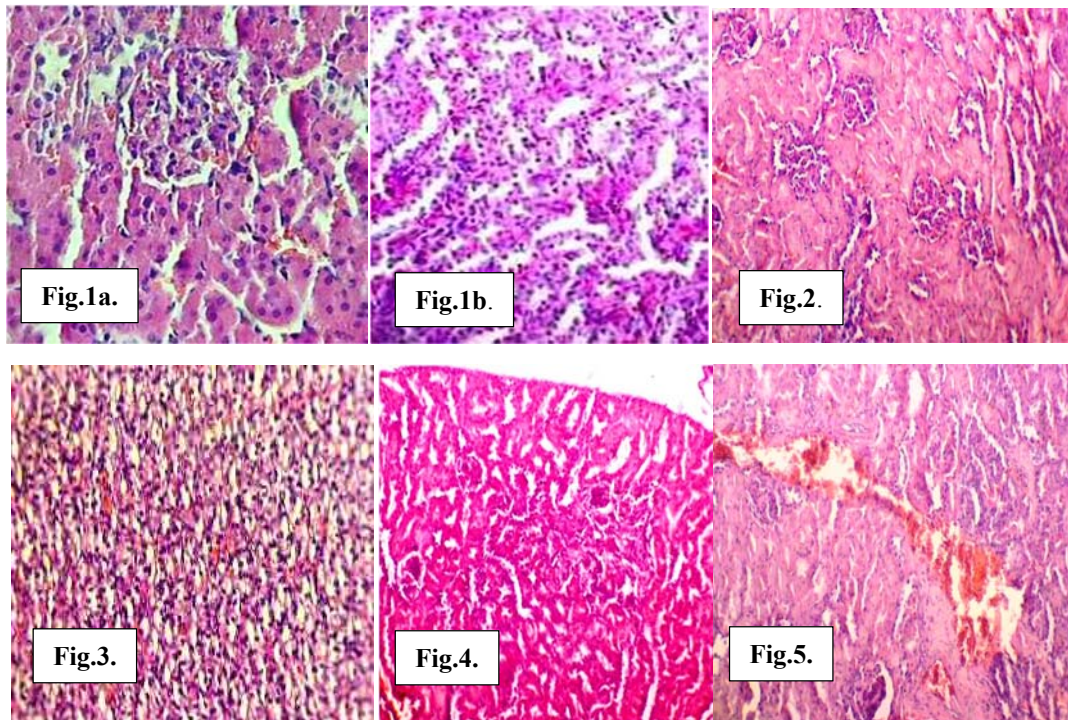


Figure 1a. A photomicrograph of a renal section of control rat (with intake of distilled water alone) .Normal architecture of renal glomeruli with intact Bowman's capsule .Brush bordered cuboidal epithelium lining the proximal convoluted tubules. Simple cuboidal epithelium lining the distal convoluted tubules.(H.&E.×400).**Figure 1b.**A photomicrograph of a renal section obtained from control rat with normal medullary renal tubules (H.&E.×250). **Figure 2.**A photomicrograph of a renal section of rat from imatinib group shows glomeruli with congested capillaries, decreased Bowman's space, cloudy degeneration of proximal convoluted tubules, and dilated distal renal tubules.(H.&E.×160).**Figure 3.**A photomicrograph of a renal section of rat administered with with 75mg/kg of imatinib daily for 30 days. Accumulation of chronic inflammatory cells (lymphocytes and macrophages) in the medulla, and extravasations of RBC.(H.&E.×250). **Figure 4.**A photomicrograph of a renal section of rat received 75mg/kg/day/30 days of imatinib with feature of early fibrosis (periglomerular and peritubular), and glomerular atrophy.(H.&E.×250). **Figure 5.**A photomicrograph of a renal section of rat received 75mg/kg/day/30 days of imatinib with dilated and congested blood vessel and glomerular shrinkage with appearance of structureless eosinophilic area of necrosis.(H.&E.×250).

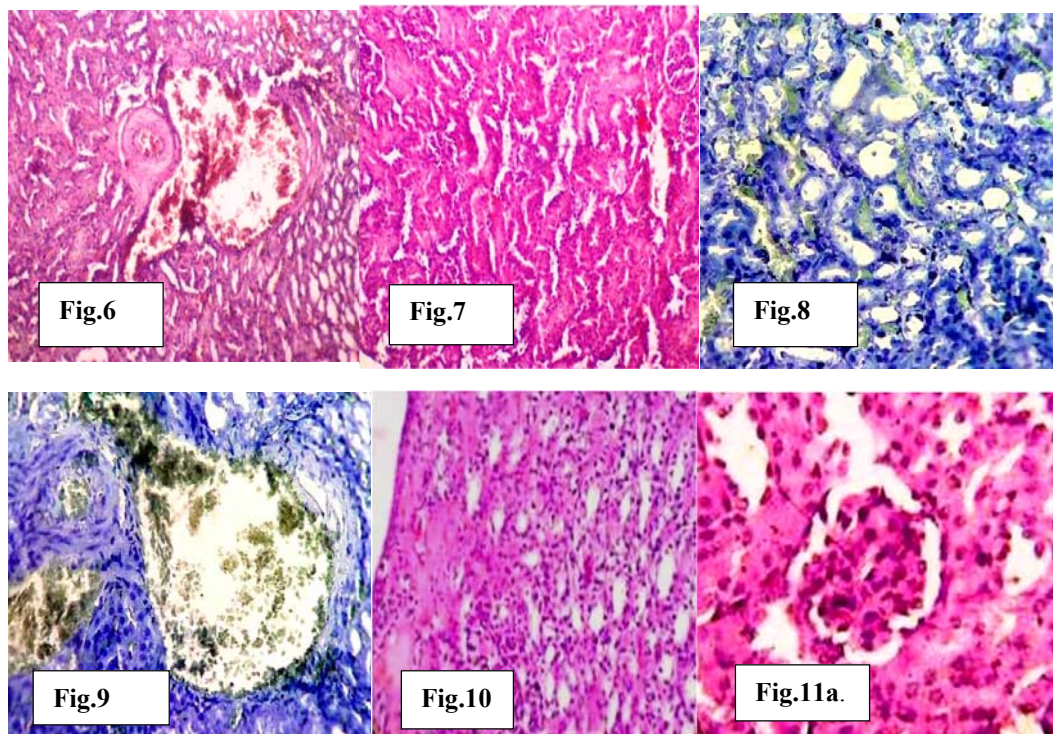


Figure 6 .A photomicrograph of a renal section of rat administered with imatinib. Dilated juxtamedullary blood vessels, thickening and hyalinization of the blood vessel wall and cloudy degeneration (H.&E.×250).**Figure 7**.A photomicrograph of a renal section of rat from imatinib group. Dilated tubules with accumulation of structureless eosinophilic homogenous material in tubular lumen, and dilated blood vessel .(H.&E.×250). **Figure 8**.A photomicrograph of a renal section of rat treated with imatinib. Dilated tubules with flattened epithelium , presence of sloughed cells in their lumina, and extravasated RBC.(Toluidine blue×250).**Figure 9**.A photomicrograph of a renal section of rat treated with imatinib with thickening of the blood vessel wall. (Toluidine blue×400).**Figure 10**.A photomicrograph of a renal section of rat treated with imatinib with evidence of interstitial oedema (homogenous area)and early inflammation.(H.&E.× 250). **Figure 11a**.A photomicrograph of sections from control rats. The normal amount of carbohydrates in cortex (PAS+H×400).

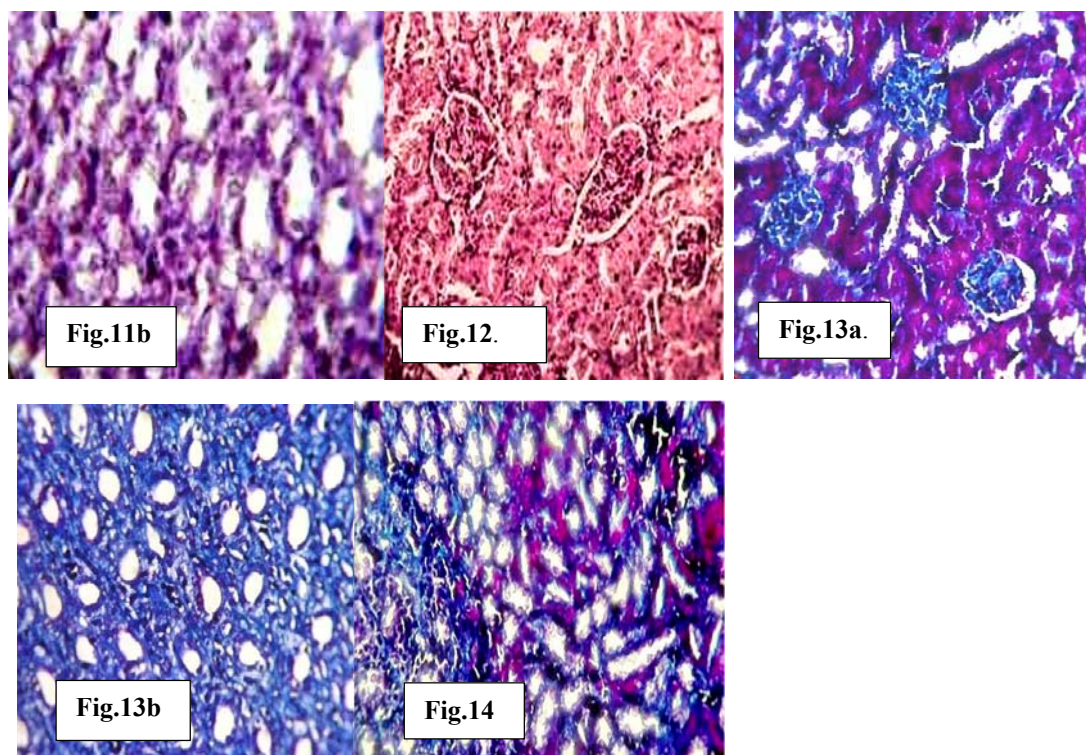


Figure 11b. A photomicrograph of sections from control rats. The normal amount of carbohydrates in medulla (PAS+H× 250). **Figure 12.** A photomicrograph of section from imatinib group. Decrease of total carbohydrates (PAS+H×250). **Figure 13a.** A photomicrograph of cortical area of control group. (Masson's Trichrome×250) **Figure 13b.** A photomicrograph of medullary area of control group. (Masson's Trichrome×250) **Figure 14.** A photomicrograph of renal section of imatinib group with cloudy appearance, focal infiltration with inflammatory cells. (Masson's Trichrome×400).

References

1. Shah RR, Morganroth J, Shah DR. Hepatotoxicity of tyrosine kinase inhibitors: clinical and regulatory perspectives. *Drug Saf.* 2013;36(7):491-503.
2. Pou M, Saval N, Vera M, Saurina A, Solé M, Cervantes F, Botey A. Acute renal failure secondary to imatinib mesylate treatment in chronic myeloid leukemia. *Leuk Lymphoma.* 2003;44: 1239-1241.
3. Fabio P S Santos, Hagop Kantarjian, Alfonso Quintás-Cardama, and Jorge Cortes. Evolution of Therapies for Chronic Myelogenous Leukemia *Cancer J.* 2011; 17(6): 465–476.
4. Gishizky ML, Johnson-White J, Witte ON. Efficient transplantation of BCR-ABL-induced chronic myelogenous leukemia-like syndrome in mice. *Proc Natl Acad Sci U S A.* 1993; 90(8):3755–3759.
5. Druker BJ, Guilhot F, O'Brien SG, et al. Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. *N Engl J Med.* 2006; 355(23):2408–2417.
6. El-Sayyad H., Ismail M.F., Shalaby F.M et al. Histopathological effects of cisplatin, doxorubicin and 5-fluorouracil (5-FU) on the liver of male albino rats. *Intern J of Biol Sci.* 2009; 5(5):466-473.
7. Deininger M, Buchdunger E, Druker BJ. The development of imatinib as a therapeutic agent for chronic myeloid leukaemia. *Blood*

- 2005;105:2640–2653.
8. Dagher R, Cohen M, Williams G. et al. Approval summary: imatinib mesylate in the treatment of metastatic and/or unresectable malignant gastrointestinal stromal tumours. *Clin Can Res.* 2002; 8:3034–3038.
 9. Kingham TP, Dematteo RP. Multidisciplinary treatment of gastrointestinal stromal tumors. *Surg Clin North Am* 2009; 89:217–233.
 10. Demetri GD, Wang Y, Wehrle E, Racine A, Nikolova Z, Blanke CD, et al. Imatinib plasma levels are correlated with clinical benefit in patients with unresectable/metastatic gastrointestinal stromal tumors. *J clin Oncol* 2009;27:3141–3147.
 11. Buchdunger E, Cioffi CL, Law N. et al. Abl protein tyrosine kinase inhibitor ST1571 inhibits in vitro signal transduction mediated by c-kit and platelet derived growth factor receptors. *J Pharmacol Exp Ther.* 2000; 295: 139-145.
 12. Handolias D, McArthur GA. Imatinib as effective therapy for dermatofibrosarcoma protuberans: proof of concept of the autocrine hypothesis for cancer. *Future Oncol.* 2008; 4: 211-217.
 13. Haberler C, Gelpi E, Marosi C. et al. Immunohistochemical analysis of platelet-derived growth factor receptor- α , - β , c-kit, c-abl, and arg proteins in glioblastoma: possible implications for patient selection for imatinib mesylate therapy. *J Neuro oncol.* 2006; 76: 105-109.
 14. Leder C, Ortler S, Seggewiss R, Einsele H, Wiendl H. Modulation of T-effect or function by imatinib at the level of cytokine secretion. *Exp Hematol.* 2007;35: 1266-1271.
 15. Lassila M, Allen TJ, Cao Z. et al. Imatinib attenuates diabetes-associated atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2004; 24: 935-942.
 16. Gschwind HP, Pfaar U, Waldmeier F, et al. Metabolism and disposition of imatinib in healthy volunteers. *Drug Metab Dispos.* 2005; 33: 1503-1512.
 17. Ramalingam S, Lagattuta TF, Egorin MJ, et al. Biliary excretion of imatinib mesylate and its metabolite CGP 74588 in humans. *Pharmacotherapy.* 2004; 24: 1232-1235.
 18. Peng B, Lloyd P, Schran H. Clinical pharmacokinetics of imatinib. *Clin Pharmacokinet.* 2005; 44: 879-894.
 19. Pou M, Saval N, Vera M, Saurina A, Solé M, Cervantes F, Botey A. Acute renal failure secondary to imatinib mesylate treatment in chronic myeloid leukemia. *Leuk Lymphoma.* 2003; 44: 1239-1241.
 20. Foringer JR, Verami RR, Tjia VM, et al. Acute renal failure to imatinib mesylate treatment in prostate cancer. *Ann Pharmacother.* 2005; 39: 2136-2138.
 21. Vickers AE, Rose K, Fisher R, Saulnier M, Sahota P, Bentley P. Kidney slices of human and rat to characterize Cisplatin-induced injury on cellular pathways and morphology. *Toxicol. Pathol.* 2004;32: 577-590.
 22. Gafter-Gvili A, Ram R, Gafter U et al. Renal failure associated with tyrosine kinase inhibitors case report and review of the literature. *Leuk Res* 2010; 34: 123–127.
 23. El-Sheikh A. A. K. Morsy MA, Mahmoud MM, Rifaai RA, Abdelrahman AM. Effect of Coenzyme-Q10 on Doxorubicin-Induced Nephrotoxicity in Rats. *Advances in Pharmacological Sciences* Article ID 2012;981461, 8 pages
 24. Soares AA, Eyff TF, Campani RB et al. Glomerular filtration rate measurement and prediction equations. *Clin Chem Lab Med* 2009; 47: 1023–1032.
 25. Marcolino MS, Boersma E, Clementino NCD, Macedo AV, Marx-Neto AD, Silva MHCR, Gelder T, Akkerhuis KM, and A. L. Ribeiro AL. Imatinib treatment duration is related to

- decreased estimated glomerular filtration rate in chronic myeloid leukemia patients. *Annals of Oncology* 2011; doi:10.1093/annonc/mdq715 | 1-7.
26. Pannabecker, T. L.; Abbott, D. E. & Dantzler, W. H. . Three dimensional Functional reconstruction of inner medullar thin limbs of Henle's loop. *Am. J. Physiol. Renal Physiol.*, 2004;286: 38-45.
 27. Al-Samawy E R M. Morphological and Histological study of the kidneys on the Albino rats Al-Anbar J. Vet. 2012; Sci.5 (1):115-119.
 28. Trevisan A , Nicolli A & Chiara F Are rats the appropriate experimental model to understand age-related renal drug metabolism and toxicity? *Expert Opin Drug Metab Toxicol.* 2010;6 (12):Pages 1451-1459 .
 29. Morgan SJ , Elangbam CS, Berens S , Janovitz E , Vitsky A , Zabka T, Conour L. Use of Animal Models of Human Disease for Nonclinical Safety Assessment of Novel Pharmaceuticals *Toxicol Pathol* 2013; 41 (3): 508-518.
 30. Favareto A.P.A, Fernandez C.D.B, Fossato da Silva D.A., Janete Aparecida Anselmo-Franci J.A., Kempinas W.D.G.. Persistent Impairment of Testicular Histology and Sperm Motility in Adult Rats Treated with Cisplatin at Peri-Puberty .*Basic & Clinical Pharmacology & Toxicology.* 2011;109:85–96
 31. Ilbey YO, Ozbek E, Cekmen M, Simsek A, Otuncemur A, Somay A. Protective effect of curcumin in cisplatin-induced oxidative injury in rat testis: mitogen-activated protein kinase and nuclear factor-kappa B signaling pathways. *Human Reproduction*, 2009.24(7): 1717–1725.
 32. Urban J.A.D'Souza. Toxic effects of 5-fluorouracil on sperm count in wistar rats .*Malaysian Journal of Medical Sciences.* 2003;10(1):43-45.
 33. Mohan M, Bhandare S .Protective effect of solanum torvum against testicular toxicity in male wistar rats. *International Journal of Pharmacy and Pharmaceutical Sciences* . 2012;4(3): 188-192.
 34. Kerkela R., Grazette L., Yacobi R., Iliescu C., Patten R., Beahm C., Walters B., Shevtsov S., Pesant S., Clubb FJ., Rosenzweig A., Salomon RN., Van Etten RA., Alroy J., Durand JB, Force T., Cardiotoxicity of the cancer therapeutic agent imatinib mesylate. *Nature Medicine* . 2006;12 (8) :908-916.
 35. Bachmann K, Pardoe D, White D. Scaling basic toxicokinetic parameters from rat to man. *Environ Health Perspect* 1996;104:400–407.
 36. Yaghmaei P., Parivar K., Gharibi A., Nabuini M. The biological effects of imatinib on male fertility of wistar rats. *International Journal of Fertility and Sterility.* 2009;3(3):135-142.
 37. Saad SY, Alkharfy KM ,and Arafah MM. Cardiotoxic effects of arsenic trioxide/imatinib mesilate combination in rats. *JPP* 2006, 58: 1–7
 38. Afify M. , Mohamed Diaa El-dien Abd Elmaksoud, Tamer Mosa, Marwa Elshaer, Nahla Kotb. Differential effects of amitriptyline treatment on testicular and liver functions in adult male rats *New York Science Journal* 2010;3(3)
 39. Nassar I, Pasupati Th., Judson JP., Segara I. Histopathological study of the hepatic and renal toxicity associated with the co-administration of Imatinib and Acetaminophen in a preclinical mouse model. *Malaysian J Pathol* .2010; 32(1):1– 11.
 40. Kose E., Sapmaz H.I., Sarihan E., Vardi N., Turkoz Y., Ekinici N. Beneficial Effects of Montelukast Against Methotrexate-Induced Liver Toxicity: A Biochemical and Histological Study. *The Scientific World Journal*. Volume 12, Article ID 987508, 6 pages. doi:10.1100/2012/987508.
 41. Thariat J, Barriere J, Janus N, Launay Vacher V (2011) Renal toxicity of targeted therapies.

- Bull Cancer 2011.
42. Steegmann JL, Cervantes F, le Coutre P, Porkka K, and Saglio G. Off-target effects of BCR–ABL1 inhibitors and their potential long-term implications in patients with chronic myeloid leukemia, 2012; Vol. 53, No. 12, Pages 2351-2361
 43. Alwin HL Loh, Arthur H Cohen. Drug-induced Kidney Disease – Pathology and Current Concepts Ann Acad Med Singapore 2009; 38: 240- 250
 44. Deveci E., Söker S., Baran Ö., Tunik S., Ayaz E. and Deveci S.() Ultrastructural changes in the kidney cortex of rats treated with lead acetate. Int. J. Morpho. 2011;29(3):1058-1061
 45. Abdul-Rahman FT, Al-Allaf LIK, Al-Nuaimy HA) The histological changes of albino mice embryos' kidneys after exposure in utero to topiramate J Med J.2011 ; Vol. 45 (3):245- 254)
 46. Pinder EM, Atwal GS, Ayantunde AA et al. Tumour lysis syndrome occurring in a patient with metastatic gastrointestinal stromal tumour treated with glivec (imatinib mesylate, Gleevec, STI571). Sarcoma 2007; 82012.
 47. Al-Kali A, Farooq S, Tfayli A. Tumor lysis syndrome after starting treatment with Gleevec in a patient with chronic myelogenous leukemia. J Clin Pharm Ther 2009; 34: 607–610.
 48. Padmini MP, Kumar JV. ().An experimental study of biochemical and histopathological study on gentamycin induced renal failure in albino rat and the effectiveness of punarnava (boerhaavia diffusa) on reversal of renal damage. IOSR Journal of Dental and Medical Sciences (IOSR-JDMS) 2013; Volume 9, Issue 6 PP 17-21.
 49. Pallotti MC, Pantaleo MA, Nannini M, Centofanti F, Fabbri B, Montanari M, Baraldi O, Saponara M, Lolli C, Mandrioli A, Biasco G. Development of a Nephrotic Syndrome in a Patient with Gastrointestinal Stromal Tumor during a Long-Time Treatment with Sunitinib. Case Rep Oncol. 2012;5(3):651-656.
 50. Baek SH, Kim H, Lee J, Kim DK, Oh KH, Kim YS, Han JS, Kim TM, Lee SH, and Joo KW. Renal adverse effects of sunitinib and its clinical significance: a single-center experience in Korea. Korean J Intern Med. 2014;29(1): 40–48.
 51. Naughton CA. Drug-induced nephrotoxicity. Am Fam Physician 2008; 78: 743–750.
 52. Floege J, Eitner F, and Alpers CE (2008) A new look at platelet-derived growth factor in renal disease. J Am Soc Nephrol 19: 12–23,
 53. Torres VE and Leof EB. Fibrosis, regeneration, and aging: playing chess with evolution. J Am Soc Nephrol 2011;22: , doi: 10.1681/ASN.2011060603
 54. Lim AYL, Segarra I, Chakravarthi S, Akram S, Judson JP. Histopathology and biochemistry analysis of the interaction between sunitinib and paracetamol in mice. BMC Pharmacology, 2010; 10:14.
 55. Carvalho C, Santos RX, Cardoso S. et al., Doxorubicin: the good, the bad and the ugly effect. Current Medicinal Chemistry, 2009; vol. 16, no. 25, pp. 3267–3285,.
 56. Hassan NAM, Yousef MM. Study of imatinib cardiotoxicity in adult male rabbits. Journal of environmental science, Toxicology And Food Technology 2013 (IOSR-JESTFT) 6(5) : 14-26
 57. Hu W, Lu S, McAlpine I, Jamieson J D, Lee DU, Marroquin LD, Heyen JR, Jessen BA. Mechanistic Investigation of Imatinib-Induced Cardiac Toxicity and the Involvement of c-Abl Kinase. Toxicol. 2012; Sci. 129 (1): 188-199.
 58. Cui J, Shi S, Sun X, Cai G, Cui S, et al. Mitochondrial autophagy involving renal injury and aging is modulated by caloric intake in aged

- rat kidneys. PLoS ONE 2013; 8(7):e69720.doi:10.1371/journal.pone.0069720
59. Altunkaynak ME, Özbek E, Altunkaynak BZ, Can I, Unal D, and Unal B. The effects of high-fat diet on the renal structure and morphometric parameters of kidneys in rats J Anat. Jun 2008; 212(6): 845–852.
60. Sakr S., Okdah A, and El-Abed F. (2003): Gibberellin A₃ induced histological and histochemical alterations in the liver of albino rats, ScienceAsia 2003;29: 327-331.

تأثير المعاملة بعقار الايماتنب على نسيج الكلى لدى الجرذان البيضاء

لمى ابراهيم خليل العلاف، حافظ علي محمود العشو *

* استاذ مساعد، فرع التشريح، كلية الطب، جامعة الموصل، الموصل، العراق

الملخص

الخلفية: ان الادوية المستهدفة للجزيمات الصغيرة قد احدثت ثورة في علاج مرض ابيضاض الدم المزمن خلال العقد المنصرم مع ان استعمال تلك الادوية قد وجد انه مرتبط بتأثيرات سمية خطيرة على عدد من الاعضاء الحيوية ومن ضمنها الكليتين.

أهداف الدراسة: تهدف الدراسة الى تحديد التغيرات النسيجية الحاصلة في كلية الجرذان المهقاة جراء التجريع بالايمانتب ميسيليت وبجرعة تساوى 75 ملغرام لكل كيلوغرام من وزنهم مرة واحدة باليوم ولمدة شهر بالمقارنة مع مجموعة السيطرة.

مكان الدراسة: هذه الدراسة التجريبية شملت تجريع ست عشر من الجرذان المهقاة والمهداة من بيتي الحيوانات التابع لكلية الطب البيطري، جامعة الموصل في مدينة الموصل شمالي العراق.

طرق العمل: تضمن العمل تجريع ثمانية من الجرذان بعمر 40-45 يوما عن طريق الفم يوميا بمقدار 75 ملغرام لكل كيلوغرام من وزن الجسم من عقار الايماتنب ميسيليت (كليفك، نوفارتس) ولمدة شهر مع مجموعة سيطرة وبنفس العمر ونفس العدد تم تجريعهم بالماء المقطر. تم اخذ الكليتين وطمرهما بالبارافين وصبغهما بعد ذلك باهيماتوكسلين ايسوين، صبغة حامض البريودك - شيف + هيماتوكسيلين، تولدين بلو، والماسون تراكروم .

النتائج: شوهد حدوث تغيرات نسيجية مختلفة في مجموعة الجرذان التي استلمت الايماتنب عند جرعة 75 ملغرام لكل كيلوغرام يوميا ولمرة واحدة لمدة ثلاثين يوما في الكبيبة وبعض اجزاء النبيبات بالمقارنة مع تلك التابعة لمجموعة السيطرة. كانت اكثر الواجه حدة هي التوسع في فراغ بومان، وجود الكبيبة المفصصة او المقسمة والمنكمشة مع حدوث توسع في النبيبات الكلوية مع التقشر في الخلايا الطلائية لهم مع وجود الاضمحلال المتبلد والمكفهر. ولوحظ ايضا احتقان في الاوعية الشعرية في الكبيبة في الشرائح النسيجية التابعة لمجموعة الجرذان المعاملة بالعقار مع حدوث ضيق في فراغ بومان. ان اذى الكلى الملحوظ والملفت كان في انتفاخ الخلايا التابعة للنبيبات الكلوية مع فقدان الحافة الفرشائية في النيبب الداني الملفوف مع وجود الاضمحلال المتبلد والمكفهر للنبيبات وكان هناك تراكم بؤرى للخلايا الالتهابية للالتهاب المبكر وقد ترشحت بين النبيبات في منطقة القشرة واللب مع حدوث تلف مبكر. ان اشرايح النسيجية التابعة لمجموعة الجرذان المعاملة بالعقار شهدت توسع في الاوعية الدموية ما بين النبيبات في منطقة القشرة واللب مع ظهور منطقة محبة للايوسين منعدمة التركيب تدل على حدوث نخر مع توسع النبيبات مع تراكم مواد متجانسة في وعاء النيبب. شهدت المنطقة البينية وجود زيادة خلوية ووذمة وارتشاح خلايا التهابية احادية النواة (لمفية) قد تركزت حول النبيبات في القشرة واللب.

الاستنتاجات: ان عقار الايماتنب قد سبب في تأثيرات سلبية على نسيجية الكلى وحدوث تحويرات في الكبيبة الكلوية وخلاياها وكذلك في نبيبات الكلى مما قد يلعب دورا مهما في خلل وظائف الكلى. ان التعاون السريري بين اخصائي الاورام واخصائي امراض الكلى قد يكون مهما لضبط الجرعة بالنسبة لمثبطات التايروسين كاينيز.

الكلمات الدالة: الايماتنب، ميسيليت، ابيضاض الدم اللوكيمي المزمن، سمية الكلى، الجرذان المهقاة.