

Analysis and Evaluation of Hydroxytyrosol in Olive Leaf Extract

Muhammed Alzweiri^{1✉} and Yusuf M. Al-Hiari¹

¹ Department of Pharmaceutical Sciences, Faculty of Pharmacy, the University of Jordan, Amman-11942, Jordan

ABSTRACT

There is a group of phenolic compounds and their glycosidic forms responsible of antioxidant effect in the olive leaf extract. Hydroxytyrosol is one of these compounds considered as building unit for the other phenolic molecules in the extract. A normal phase chromatographic method was established for analysis of hydroxytyrosol. It generates clean chromatograms suitable for analytical and preparative purposes and better resolved than those obtained from reversed phase systems. Mobile phase of (1:1) acetonitrile and 1% acetic acid aqueous solution was used. Flow rate was kept constant at 0.5 mL/min. Injection volume was 20 μ L and UV detector was set at $\lambda=280$ nm. The developed HPLC method was found linear within the range of 0.82-4.12 mg%, precise with RSD less than 2% and accurate with a range of 97.6-101.2 %. LOQ and LOD of the method were calculated to be 8 and 0.8 μ g/ml respectively. Furthermore, antioxidant degree of hydroxytyrosol and its derivatives in the extract was evaluated by Follin-ciocalteu's reagent and found equivalent to 40 mg gallic acid.

Keywords: Hydroxytyrosol, Olive leaf and Antioxidant.

INTRODUCTION

Olive leaf has a reported use in preserving food and also it has strong potential to be used in pharmaceutical preparations¹⁻³. Leaf extract is used in treatment of diabetes and cardiovascular diseases⁴⁻⁷. The activity of extracts could be attributed to phenolic constituents which possess antimicrobial and antioxidant effects^{8,9}.

According to the literature, olive leaf extract contains group of antioxidant compounds¹⁰⁻¹². Hydroxytyrosol and Oleuropein are the major ones **Figure.1**¹³⁻¹⁵. Hydroxytyrosol is the primary structure of antioxidants in olive leaf which might be further modified and glycosylated to oleuropein and other glycosides^{16,17}.

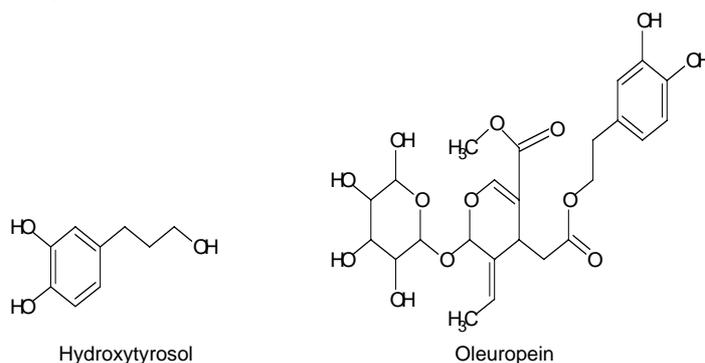


Figure.1 Structure of major metabolites in olive leaves; Hydroxytyrosol and oleuropein.

✉ m.alzweiri@ju.edu.jo

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Antioxidants are quite susceptible to decomposition due to heat and light¹⁸⁻²⁰. Thus, the total content of phenolic compounds in the extract are analysed by non-destructive test such as Folin-Ciocalteu phenol test²¹⁻²³. However, stability-indicating method is also essential to periodically evaluate the stability of products containing the plant extract²⁴. Since hydroxytyrosol is one of major compounds in the extract and it is responsible of part of antioxidant activity, it is used for standardizing the extract relying on an HPLC-dependent method^{24,25}. Hydroxytyrosol analysis was found challenging because of its amphiphilic nature^{26,27}.

However, several chromatographic separations of hydroxytyrosol from different reversed phase systems were reported²⁸⁻³⁰. Separation of hydroxytyrosol from these systems is often required gradient mobile phase. The latter often crumbles the stability of chromatographic systems and the repeatability of results^{28,29}. Additionally, the published work on simple isocratic systems with reversed phase column generates weakly resolved peaks for hydroxytyrosol³⁰. Even if resolution is enough for analytical purposes, it might not be satisfactory for preparative application by which hydroxytyrosol can be isolated.

This work introduced a new normal phase chromatographic method obtained the advantage of amphiphilic character of hydroxytyrosol in chromatographic separation. Clean chromatograms obtained from the method and the system stability due to isocratic flow enable accurate standardization of hydroxytyrosol. Also clean and well-resolved chromatograms facilitate the application of the method in preparative analysis to produce hydroxytyrosol in pure form and with cheap cost. This is expected to generate positive impact on countries relying on olive tree as one of their economic resources including pharmaceutical and food industries.

Material and methods

Reagents and chemicals

All chemicals were purchased from Sigma- Aldrich (St. Louis, MO) unless otherwise indicated. Also all the

organic solvents were obtained from Fisher Scientifics, Pittsburgh, PA, USA

HPLC standardization of Olive extract according to the level of hydroxytyrosol.

Preparation of working standard

0.5Kg of fresh leaves were collected from the campus of The University of Jordan and taxonomically identified. Then they were cut and soaked in 500ml methanol to enhance the extractability of hydroxytyrosol. The mixture left for 24 hours with frequent stirring. The extract was completely evaporated and the residue was reconstituted with 200ml ether. Subsequently, the solution was filtered to remove ether-insoluble compounds and evaporated. 100ml of water was added to reconstitute the hydroxytyrosol residue which possessing amphiphilic solubility character. The identity of hydroxytyrosol was confirmed by several systems of HPLC-UV against a relatively-expensive standard material of hydroxytyrosol (phytoplan GmbH, Heidelberg). The standardized working standard solution was entirely dried by freeze drying technique (lyophilization) and stored in refrigerator until used.

Preparation of samples

Methanolic extract of olive leaves were filtered to remove the generated insoluble particles. The filtrate was dried completely and the residue was then reconstituted with 20ml ether. After that the solution was filtered to remove ether-insoluble compounds and then evaporated. 1ml of water was added to reconstitute the residue. The resulted solution was used to quantify the amount of hydroxytyrosol in extract samples against the working standard.

Chromatographic conditions

HPLC Instrument (Shimadzu, Japan) equipped with a LC-20AT Pump, and SPD-20A UV detector. The chromatographic experiments were performed on a normal phase column (Hypersil silica column 25X4.6cm, 5µm). Mobile phases were applied with different ratios of acetonitrile and 1% acetic acid aqueous solution namely, 20, 50, 80 % acetonitrile for standardizing and confirming the identity of hydroxytyrosol working

standard whereas 50% acetonitrile was selected to monitor the sample stability against the working standard. Flow rate was kept constant at 0.5 mL/min. Injection volume was 20 μ L and UV detector was set at $\lambda=280$ nm. Additionally, outcomes of this normal phase system were compared with reversed phase system applied under the same conditions by using of C18 column (Ultrasphere, 25X4.6cm, 5 μ m).

Evaluation of total phenolic content in the olive extract by using Follin-ciocalteu's reagent.

Preparation of the sodium carbonate reagent

200 g of anhydrous sodium carbonate (Laboratory Rasayan, India) was dissolved in 800ml water and heated to boil then left for 24 hours at room temperature after that the solution was filtered and the volume completed to one liter.

Test procedure

14g of fresh olive leaves were cut and soaked in 100ml methanol for 24 hours with frequent shaking. 1ml of it was further diluted to 100ml with methanol. 0.5ml of the final solution was added to 8ml of distilled water and 0.5ml of Follin-ciocalteu's phenol reagent, 2N. The mixture was shaken promptly (<8min). 1.5 ml of sodium carbonate reagent was added and the samples were incubated for 2 hours. Blank preparation was carried out with the same procedure with using methanol instead of extract. Additionally, group of gallic acid solutions treated with same manner were used to establish calibration curve suitable for calculating gallic acid equivalent of the total phenols in the extract. Final concentrations of gallic acid solutions were 0.06425, 0.1285, 0.257, 0.514 and 1.285mg%. Absorbances of solutions were measured by UV-spectrophotometer. The

background absorbance (extract without treatment) has no light absorption within the region of 500-800 nm.

UV-spectrophotometer

All spectroscopic measurements were undertaken using a Spectro UV-VIS Double Beam PC Scanning Spectrophotometer (Model UVD-2950, Labomed , Inc., USA). The analysis was carried out at $\lambda=760$ nm.

Results and discussion

There is a group of phenolic compounds and their glycosidic forms responsible of antimicrobial effect in the olive leaf extract. Thus it has been decided to evaluate the content of the total phenols by colorimetric assay method. Meanwhile, it is necessary to also establish a chromatographic method to standardize olive leaf extract. Because hydroxytyrosol is one of the major compounds in olive leaf extract and also it is considered as building unit for the other phenolic molecules in olive leaves, it was decided to use it in standardization of the extract.

HPLC standardization of the extract according to the level of hydroxytyrosol.

The reference standard of hydroxytyrosol was tested depending on reversed phase chromatography with several ratios of water and acetonitrile as mobile phase. It was found that the retention and elution of hydroxytyrosol is not satisfactory enough. Fortunately, hydroxytyrosol is amphiphilic compound so it might be better retained in a normal phase column. Thus a normal phase chromatography using silica column was established for hydroxytyrosol, the results show a well retained peak for hydroxytyrosol promising to pass the validation requirements as shown in **Figure 2, A**.

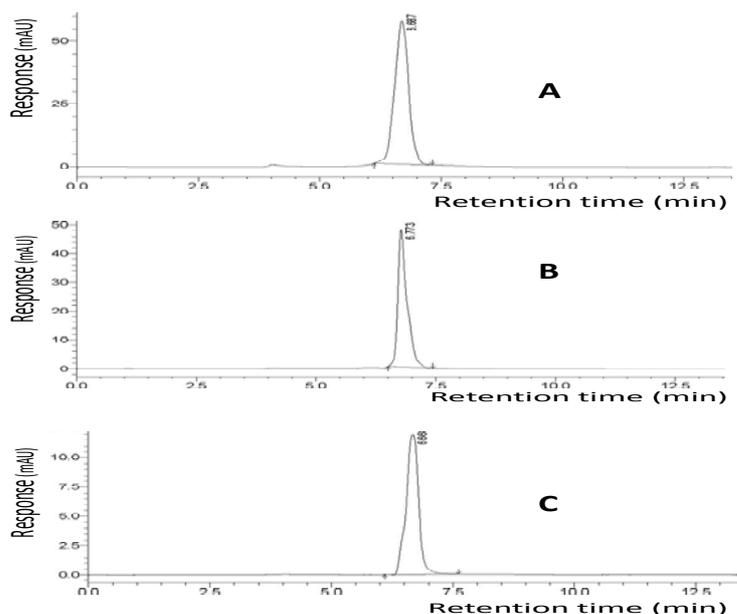


Figure 2 Chromatograms from a normal phase HPLC system for hydroxytyrosol reference standard (A), hydroxytyrosol working standard extracted from olive leaves (B) and hydroxytyrosol in a sample of olive leaf extract (C).

However, the reference standard is quite expensive and sometimes unavailable for routine analysis use. Thus it has been decided to extract hydroxytyrosol from the olive leaves as working standard after standardizing it with the reference standard material. The amphiphilic character of hydroxytyrosol, which gives it a good solubility in polar and non-polar solvents, enables its purification from the other compounds in the extract. **Figure 2, B** depicts a chromatogram of the working standard prepared from the olive leaf extract.

Working standard identity was confirmed by retention time similarity with the reference standard material under several compositions of the mobile phase as described in materials and methods topic. Although the working standard is not entirely pure, its content of hydroxytyrosol was determined quantitatively against a pure reference standard material and kept dried until used. HPLC analysis of extract samples depicts pure sharp peak for hydroxytyrosol as shown in **Figure 2, C**. Samples preparation helps in removing water soluble antioxidants such as oleuropein which has strong potential to overwhelm the obtained chromatogram. Additionally, chromatographic separation by normal phase system resolves hydroxytyrosol peak

from other interfering compounds along relatively long time period (*ca* 15 min). Interfering compounds were washed from the system between run times of samples. The obtained baseline resolution of hydroxytyrosol peak in a clean chromatogram enables transferring this method from analytical type to preparative one. Additionally, the applied isocratic system is able to keep better stability and precision than any gradient system. Simplicity and stability of the method as well as clean chromatogram facilitate the application of the method in preparative mode to produce hydroxytyrosol in pure form and with cheap cost. This should have a positive impact on countries relying on olive tree as one of their economic resources including pharmaceutical and food industries.

However, the vast majority of published work about chromatographic separation of hydroxytyrosol was implemented by reversed phase system²⁸⁻³⁰. It was found that baseline resolution of hydroxytyrosol under simple isocratic was rather difficult. The best obtained separation for hydroxytyrosol from reversed phase system is shown at **Figure 3**. This separation will not be satisfactory unless gradient systems are used. However, applying gradient systems make upgrading the analytical method to preparative analysis noisy and troubleshooting.

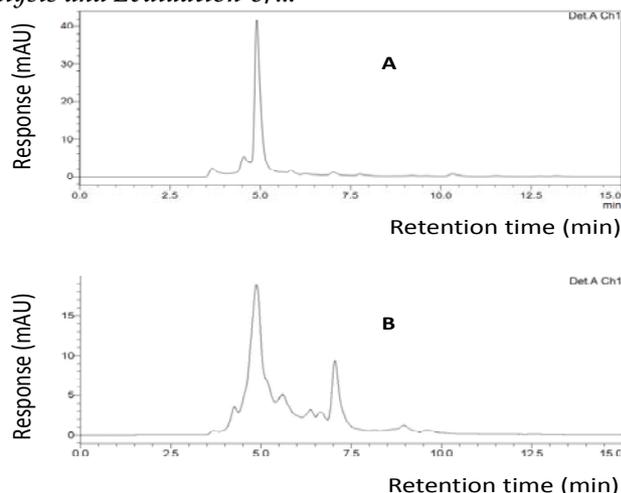


Figure.3 Chromatograms from a reversed phase HPLC system for hydroxytyrosol reference standard (A) and hydroxytyrosol in a sample of olive leaf extract (B). Baseline resolution of hydroxytyrosol was not achieved by applying a reversed phase system.

Validation of HPLC method

Linearity

Stock solution of working material equivalent to 10.3mg% of hydroxytyrosol was prepared and further diluted to six solutions with the following concentrations:

4.12, 3.29, 2.47, 2.06, 1.24 and 0.82 mg%. Each solution was injected triplicate and the average responses were plotted against concentrations as shown in **Figure 4**.

The regression coefficient shows that the method is linear over the range (0.82-4.12) mg%.

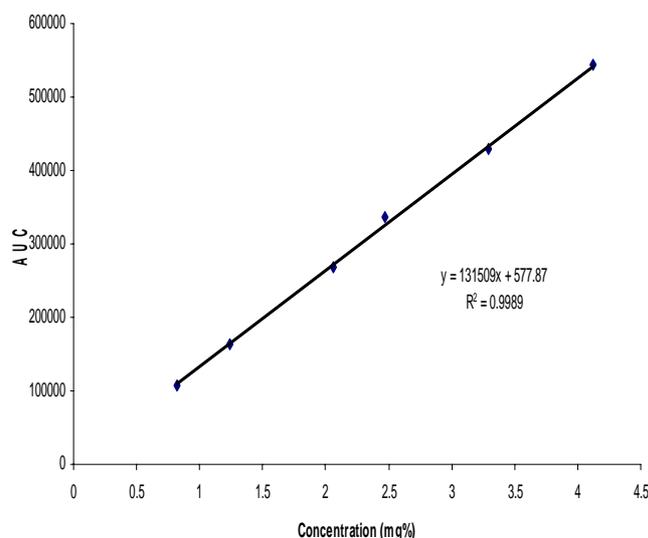


Figure. 4 linearity study of HPLC method of analysis. Response versus different concentrations of hydroxytyrosol prepared from working standard material.

Precision and Accuracy

Six solutions of working standard material with 2 mg/ml equivalent concentration of hydroxytyrosol were prepared to evaluate method precision. Moreover, other solutions of the working standard incorporated in pretested olive leaf extract were tested with the same manner to evaluate the recovery of hydroxytyrosol as

shown in **Table 1**. Triplicate injections were carried out for each solution. It was found that relative standard deviations (RSD) of inter-day and intra-day were less than 2.0% (**Table 1**). And also percentages of recovery for all prepared samples are within the range of 97.6-101.2 % (**Table 1**).

Table 1. The results of hydroxytyrosol analytical validation including accuracy and precision.

Trial#	Recovery of added 50%	Recovery of added 100%	Recovery of added 200%	Inter-day Precision (RSD)	Intra-day precision(RSD)
1	49.0±0.5	100.2±0.8	199.2±0.9	1.9%	0.7%
2	48.8±0.6	98.6±1.1	202.4±1.3	1.6%	1.5%
3	51.2±0.2	99.2±1.7	201.9±1.5	1.1%	0.6%

Limits of quantification (LOQ) and detection (LOD)

The limit of quantification (LOQ) was determined according to the least concentration below which linearity of calibration curve deviate from the norm. It was found that LOQ = 8 µg/ml. The limit of detection(LOD) was determined according to three times signal-to- noise ratio (3S/N) procedure. It was found to be 0.8 µg/ml.

Selectivity and Specificity

Selectivity of the method was assessed by comparing chromatograms of blank solvent with sample chromatograms. Specificity of the developed method was assessed by performing forced degradation study. Three solutions were prepared and heated at 80°C for 15 minutes. One of the solutions contained few drops of concentrated HCl solution. A second solution contained few drops of

H₂O₂ (30% w/v) and the third one contained NaOH (40% w/v). The purity of the hydroxytyrosol peak was tested by purity test of photodiode array detector. The purity indices of the results were above the purity threshold limit (>0.996). This confirms the purity of chromatographic peak from its degradation products. Consequently, the purity of peak from the interference of degradation products confirms the suitability of it as a stability indicating method of analysis.

Evaluation of total phenols by Follin-ciocalteu’s phenol reagent

A group of Gallic acid solutions was prepared according to the procedure mentioned in experimental section. The samples show linear curve suitable to interpolate the gallic acid equivalent value of the olive leaf extract as shown in **Figure 5**.

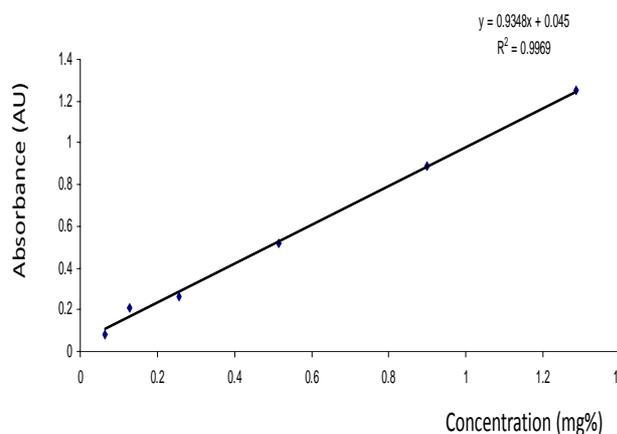


Figure 5 Linear curve of gallic acid tested according to Follin-Ciocalteu's method

On the other hand, the olive leaf extract prepared according to the method mentioned in the experimental part gave rise an absorbance value equal to 0.32. Interpolation of this value in the gallic acid curve shown at **Figure 5** resulted in gallic acid equivalence of 0.3mg%. Depending on this result, phenol content in each

1g olive leaf is equivalent to 40mg of gallic acid.

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REFERENCES

- (1) Keskin D, Ceyhan N, Ugur A, Dbeys AD. Antimicrobial activity and chemical constitutions of West Anatolian olive (*Olea europaea* L.) leaves. *Journal of Food, Agriculture & Environment*. 2012;10(2, Pt. 1):99-102.
- (2) Hudolin Kolar M, Urbancic S, Dimitrijevic D, Hojnik NM; (Vitiva d.d., Slovenia). assignee. Protection of color and anti oxidative and anti microbiological protection of meat and meat products. Application: SISI patent 2010-26023469. 2012 20100831.
- (3) Botsoglou E, Govaris A, Moulas A, Botsoglou N. Oxidative stability and microbial growth of turkey breast fillets during refrigerated storage as influenced by feed supplementation with olive leaves, oregano and/or α -tocopheryl acetate. *British Poultry Science*. 2010;51(6):760-8.
- (4) Cvjeticanin T, Miljkovic D, Stojanovic I, Dekanski D, Stosic-Grujicic S. Dried leaf extract of *Olea europaea* ameliorates islet-directed autoimmunity in mice. *British Journal of Nutrition*. 2010;103(10):1413-24.
- (5) Jemai H, El Feki A, Sayadi S. Antidiabetic and Antioxidant Effects of Hydroxytyrosol and Oleuropein from Olive Leaves in Alloxan-Diabetic Rats. *Journal of Agricultural and Food Chemistry*. 2009;57(19):8798-804.
- (6) Zrelli H, Matsuoka M, Kitazaki S, Araki M, Kusunoki M, Zarrouk M, Miyazaki H. Hydroxytyrosol Induces Proliferation and Cytoprotection against Oxidative Injury in Vascular Endothelial Cells: Role of Nrf2 Activation and HO-1 Induction. *Journal of Agricultural and Food Chemistry*. 2011;59(9):4473-82.
- (7) Poudyal H, Campbell F, Brown L. Olive leaf extract attenuates cardiac, hepatic, and metabolic changes in high carbohydrate-, high fat-fed rats. *Journal of Nutrition*. 2010;140(5):946-53.
- (8) Tsimidou MZ, Papoti VT. Bioactive ingredients in olive leaves. *Olives and Olive Oil in Health and Disease Prevention*. 2010:349-56.
- (9) Japon-Lujan R, Janeiro P, Luque de Castro MD. Solid-Liquid Transfer of Biophenols from Olive Leaves for the Enrichment of Edible Oils by a Dynamic Ultrasound-Assisted Approach. *Journal of Agricultural and Food Chemistry*. 2008;56(16):7231-5.
- (10) Valavanidis A. Antioxidant and anticancer substances of olive oil and olive leaves and their contribution to human health. *Chemika Chronika, Genike Ekdose*. 2009;71(9):25-8.
- (11) Lee O-H, Lee B-Y. Antioxidant and antimicrobial activities of individual and combined phenolics in *Olea europaea* leaf extract. *Bioresource Technology*. 2010;101(10):3751-4.
- (12) Ha JY, Goo SY, Sung J-S, Shin H-S. Antioxidant and cytoprotective activity of the olive leaf (*Olea europaea* L. var. Kalamata) extracts on the mouse embryonic fibroblast cell. *Food Science and Biotechnology*. 2009;18(4):965-70.
- (13) Ye J-z, Wang C-z, Chen H-x, Dong Y-h. Content determination and extraction technology of hydroxytyrosol from olive leaves. *Linchan Huaxue Yu Gongye*. 2011;31(1):63-7.
- (14) Fu S, Wan W, Yang W, Hu Y. Separation and purification of oleuropein from olive leaves. *Shengwu Jiagong Guocheng*. 2011;9(4):31-4.
- (15) Uematsu K, Matsui K, Shibazaki H, Hayakawa S, Ogawa M; (Kagawa Prefecture, Japan; Kagawa University; Yamahisa K. K.). assignee. Olive leaf extracts with high oleuropein content, their manufacture using citric acid, and masking agents for them. Application: JPJP patent 2009-2890782011125301. 2011 20091221.
- (16) Bu W, Liu C, Tian S. Comparison of extraction of olive leaves to prepare hydroxytyrosol by hydrochloric acid and α -glycosidase. *Shipin Gongye Keji*. 2011;32(7):228-32.
- (17) Di Donna L, Mazzotti F, Salerno R, Tagarelli A, Taverna D, Sindona G. Characterization of new phenolic compounds from leaves of *Olea europaea* L. by high-resolution tandem mass spectrometry. *Rapid Communications in Mass Spectrometry*. 2007;21(22):3653-7.
- (18) Hayashi N, Yamamoto K; (Fujifilm Corporation, Japan). assignee. Evaporable or thermally decomposable antioxidant-containing coating composition for fabricating charge transport and

- electroluminescent layers of organic electroluminescent device. Application: WOWO patent 2011-JP724822012043774. 2012 20110929.
- (19) Lin Q, Pan L, Fu S, Xu D, Wang G, Lu L, Chen K, Liu H, Wang L, Pang S, Xu J, Cao Y, Chen B; (China National Offshore Oil Corp., Peop. Rep. China; China Blue Chemical Ltd.; Hainan University; CNOOC Green Materials Co., Ltd.). assignee. Biodegradable polypropylene carbonate composite and its preparation method. Application: CNCN patent 2009-10089729101962470. 2011 20090722.
- (20) Yamashita H, Kawaguchi AW, Ohkatsu Y. New antagonism of hindered amine light stabilizers with acidic compounds including phenolic antioxidants (part 2) formation mechanism of active species of peroxide decomposition reaction. *Journal of the Japan Petroleum Institute*. 2006;49(6):294-300.
- (21) Sundaram RS, Ramanathan M, Gowtham L, Jena PK, Choudhury GB, Manikandan P, Venugopal V, Kamalakannan D. Investigation of standardized ethanolic extract of *Ocimum sanctum* Linn. (holy basil) leaves for its in vitro antioxidant potential and phenolic composition. *Asian Journal of Chemistry*. 2012;24(4):1819-24.
- (22) Spiridon I, Bodirlau R, Teaca C-A. Total phenolic content and antioxidant activity of plants used in traditional Romanian herbal medicine. *Central European Journal of Biology*. 2011;6(3):388-96.
- (23) Shukla S, Saluja AK, Pandya SS. In-vitro antioxidant activity of aerial parts of *Lippia nodiflora* Rich. *Pharmacologyonline*. 2009(2):450-9.
- (24) Steinigen M. Analysis of plant drugs. Paperback APV. 1984;11(Qual. Pflanz. Arzneim.: Pruef. Herstell.--Anforderungen Zulassung):115-25.
- (25) Bouaziz M, Feki I, Ayadi M, Jemai H, Sayadi S. Stability of refined olive oil and olive-pomace oil added by phenolic compounds from olive leaves. *European Journal of Lipid Science and Technology*. 2010;112(8):894-905.
- (26) Visioli F, Colombo C, Galli C. Differential partitioning of antioxidants, including hydroxytyrosol, in human plasma and LDL: Implications for their antioxidant activity in vivo. *Food Chemistry*. 2012;132(1):499-501.
- (27) Rietjens SJ, Bast A, de Vente J, Haenen GRMM. The olive oil antioxidant hydroxytyrosol efficiently protects against the oxidative stress-induced impairment of the NO response of isolated rat aorta. *American Journal of Physiology*. 2007;292(4, Pt. 2):H1931-H6.
- (28) Rodríguez-Gutiérrez G, Wood S, Fernández-Bolaños Guzmán J, Duthie GG, de Roos B. Determination of 3,4-dihydroxyphenylglycol, hydroxytyrosol and tyrosol purified from olive oil by-products with HPLC in animal plasma and tissues. *Food Chemistry*. 2011;126(4):1948-52.
- (29) Tasioula-Margari M, Okogeri O. Simultaneous determination of phenolic compounds and tocopherols in virgin olive oil using HPLC and UV detection. *Food Chemistry*. 2001;74(3):377-83.
- (30) Sannino F, De Martino A, Capasso R, El Hadrami I. Valorisation of organic matter in olive mill wastewaters: Recovery of highly pure hydroxytyrosol. *Journal of Geochemical Exploration*. 2013;129(0):34-9.

