

Influence of Dexamethasone on Pharmacokinetic Parameters of Cyclosporine in Rabbits

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ABSTRACT

The present study was designed to investigate the presence of significant pharmacokinetic (PK) interaction between Dexamethasone (DEX) at different concentrations on Cyclosporine (CsA) PK parameters in healthy male rabbits. The rabbits were selected and divided into three groups: Control group (n=6) received orally CsA solution (7.5 mg/kg/day) prepared from soft gelatin capsule for five days and on the fifth day, serial blood samples were withdrawn from marginal ear vein of rabbits at different time intervals post-dosing. In the first and second groups, rabbits were given orally (7.5 mg/kg/day) CsA solution concomitantly with DEX at two doses (0.33 and 0.66 mg/kg/day), respectively. On the fifth day of administration, each test group's serial blood samples were collected for over 24 hours as in the control group. Different PK parameters of CsA for the three groups were determined using non-compartmental analysis. It was observed that, there were statistically insignificant differences between control and test groups when co-administered with DEX at both concentrations. The present study results demonstrated that concurrent administration of DEX at both concentrations had not influenced the PK parameters of CsA.

Keywords: cyclosporine, dexamethasone, drug-drug interaction, pharmacokinetic parameters.

INTRODUCTION

CsA is a calcineurin inhibitor and potent immunosuppressive agent, which significantly impacts organ transplantation¹. It has considerably improved the first and second-years graft survival rates and decreased morbidity in kidney, liver, heart, lung, and pancreas transplantation. In addition to that, several studies have supported the efficacy of CsA in preventing graft-versus-host disease in bone marrow transplantation². CsA is

extensively metabolized in the liver by CYP3A4 system³. Sustained and clinically significant drug-drug interactions (DDIs) can occur during long-term therapy. Thus, the co-administration of multiple drugs with CsA could increase the risk of treatment failure, nephrotoxicity, and other adverse effects⁴.

DDIs are one of the commonest causes of medication errors in developed countries, mainly in the elderly due to poly-therapy, with a prevalence of 20-40%⁵; thus, Poly-therapy increases the complexity of therapeutic management and the risk of clinically significant DDIs which can both induce the development of adverse drug

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reactions or reduce the clinical efficacy⁶. CsA soft gelatin capsule (Neoral®) is a modified micro-emulsion formulation with uniform and slightly increased bioavailability compared to (Sandimmune®)^{7,8}. After oral administration of CsA (Neoral®), the time to peak blood concentration (t_{max}) is 1 to 2 hours (h), the elimination of CsA is primarily biliary with only 6% of the dose (parent drug and metabolites) excreted in urine, the disposition of CsA from blood is generally biphasic with a terminal half-life ($t_{1/2}$) approximately 8.4 h (range 5 to 18 h)⁹. Dexamethasone (DEX) is a synthetic adrenocortical steroid agent used in treating chronic inflammatory and autoimmune diseases and is also effective as an antiemetic agent in cancer chemotherapy¹⁰. Cytochromes P450 (CYP) is a major source of variability in drug PKs, and response¹¹ and CYP3A4 is the most common and versatile one¹². Many clinically significant drug interactions result from induction or inhibition of CYP3A4 enzymes, the major drug-metabolizing enzymes mainly in the liver^{13,14}.

A remarkable feature of CYP3A4 is its extreme promiscuity in substrate specificity and cooperative substrate binding, which often leads to undesirable DDIs and toxic side effects. Owing to its importance in drug development and therapy, CYP3A4 has been the most extensively studied¹⁵. This study aims to investigate the presence of clinically significant PK interaction between DEX at different concentrations on CsA (A narrow therapeutic index drug) PK parameters by using healthy male rabbits as an animal model.

1. MATERIALS AND METHODS

2.1 Animals

Several experimental trials were performed on 18 New Zealand strains of adult male rabbits weighted (3.1-3.4 kg) and aged 8-10 months were enrolled in this study. The Research and Ethics Committee approved animals of the Experimental Animal Care Facility, College of Pharmacy, Al-Azhar University of Gaza (AUG), Palestine. The rabbits were selected randomly and divided into three groups (six for each group). All rabbits were kept under

standard laboratory conditions in a 12-hour light/dark cycle at $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$ provided with pellet diet with water *ad libitum* and were fasted overnight before the experiments.

2.2 Study design and blood sampling

In an in-vivo drug-drug interaction, a randomized designed study was conducted in eighteen healthy male rabbits. The rabbits were selected and divided into three groups: Control group (n=6) received orally CsA solution (7.5 mg/kg/day) prepared from soft gelatin capsule (Neoral®) for five days, and on the fifth-day blood samples (1.5-2 mL) were withdrawn from marginal ear vein of rabbits at 0.00, 0.50, 1.00, 1.50, 2.00, 3.00, 6.00, 12.00 and 24 hr post-dosing¹⁶. The rabbits in the first and second groups (test groups) received orally CsA solution (7.5 mg/kg/day) and DEX concomitantly at two different doses (0.33 and 0.66 mg/kg/day), respectively. On the fifth day of the administration, serial blood samples from each group were collected for over 24 hours as in the control group. Whole blood samples in EDTA tubes were kept at $(2-8)^{\circ}\text{C}$ until analyzed (Whole blood sample is stable for up to 3 days).

2.3 Analysis of blood samples

Analysis of whole blood samples to determine the concentrations of CsA was performed at the laboratory of Medical Relief Society-Gaza using Maglumi 800 System and Maglumi 800 CsA detection kit (Shenzhen New Industries Biomedical Engineering Co., Ltd.). The Kit is based on chemiluminescent immunoassay (CLIA). It is used in hospitals for rapid CsA assaying in whole blood to monitor the CsA dose.

2.4 CsA PK and statistical analysis

PK parameters for control and test groups including C_{max} , t_{max} , K_e , $t_{1/2}$, AUC_{0-24} , $AUC_{0-\infty}$, and MRT were determined. Both parameters (C_{max}) and (t_{max}) were directly determined from the plasma concentration versus time curves. The linear trapezoidal rule calculated the AUC_{0-24} . The $AUC_{0-\infty}$ was determined by the following equation: $AUC_{0-\infty} = AUC_{0-24} + Ct / k_e$, where Ct is

defined as the final measured serum concentration at time t and K_e is the elimination rate constant. The K_e was determined by the least-squares regression of plasma concentration-time data points lying in the terminal region using semilogarithmic dependence that corresponds to first-order kinetics. The $t_{1/2}$ was calculated as $0.693/K_e$. PK analysis was determined using model-independent method (Non-Compartmental Approach) WinNonlin Professional Software (Version 6.3, Pharsight Corporation, Cary, NC) and (GraphPad Prism versión 4.00; San Diego, CA, USA). Statistical methods, including descriptive analysis and Mann-Whitney test, were applied to compare the PK parameters of CsA alone (control group) or co-administered with DEX in first and second groups (test groups). (SPSS) program (version 16.0) was applied to analyze data. A statistically significant difference was

considered when $P \leq 0.05$.

2. RESULTS

The DEX-CsA interaction study was carried out to determine the influence of DEX at different concentrations on the PK parameters of CsA. Whole blood concentration-time profiles of CsA in the control group (Received only CsA 7.5 mg/kg/day) and test groups (First and second groups), which received CsA concomitantly with DEX at doses (0.33, and 0.66 mg/kg/day) respectively are shown in Figure 1. The PK parameters of CsA were determined for the control and test groups, including C_{max} , t_{max} , $t_{1/2}$, K_e , AUC_{0-24} , $AUC_{0-\infty}$, and MRT. The control group's obtained PK parameters were compared with those of the first test group (Table 1) and those with the second test group (Table 2).

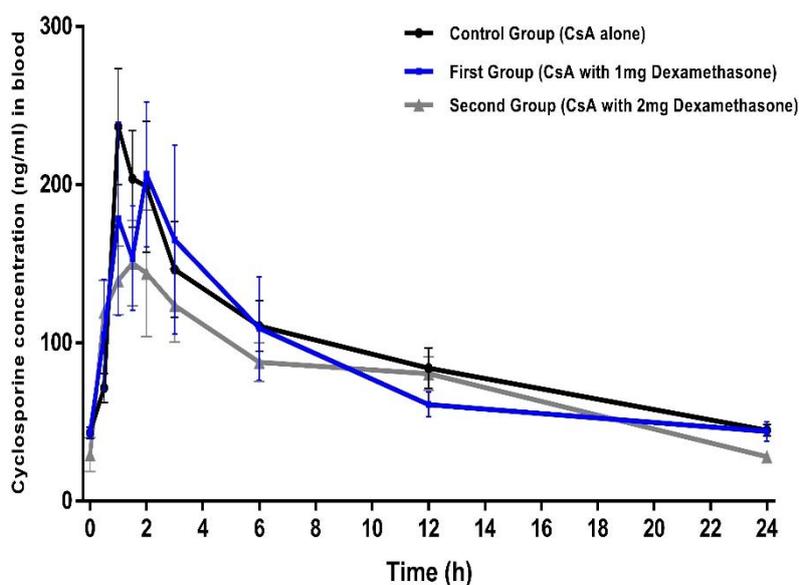


Figure 1: Plot of blood CsA concentration-time profile. Control group: Rabbits received CsA alone (7.5mg/kg/day) orally for five days, first and second groups (test groups): Rabbits received CsA (7.5mg/kg/day) concurrently with DEX (0.33 and 0.66mg/kg/day) orally for five days, respectively (n=6).

Table 1. PK parameters of CsA in control and first test group (n=6).

PK Parameters	Groups	N	Mean \pm SD	P-value
C_{max} (ng/ml)	Control group	6	254.9 \pm 106.4	0.68
	First group	6	290.0 \pm 144.9	
t_{max} (h)	Control group	6	1.50 \pm 0.87	0.58
	First group	6	1.83 \pm 0.75	
Ke (h ⁻¹)	Control group	6	0.052 \pm 0.001	0.77
	First group	6	0.054 \pm 0.02	
$t_{1/2}$ (h)	Control group	6	13.38 \pm 1.82	0.99
	First group	6	13.29 \pm 2009	
AUC ₀₋₂₄ (ng/ml.h)	Control group	6	2221 \pm 730	0.70
	First group	6	2009 \pm 946	
AUC _{0-∞} (ng/ml.h)	Control group	6	3069 \pm 808	0.71
	First group	6	2863 \pm 985	
MRT (h)	Control group	6	9.22 \pm 0.62	0.90
	First group	6	9.16 \pm 1.12	

Control group: Rabbits received CsA (7.5mg/kg/day) orally for five days; First group: Rabbits received CsA (7.5mg/kg/day) concurrently with DEX (0.33mg/kg/day) orally for five days; Statistical significance: P \leq 0.05.

Table 2. PK parameters of CsA in control and second test groups (n=6).

PK Parameters	Group	N	Mean \pm SD	P-value
C_{max} (ng/ml)	Control group	6	254.9 \pm 106.4	0.51
	Second group	6	213.5 \pm 72.6	
t_{max} (h)	Control group	6	1.50 \pm 0.87	0.58
	Second group	6	1.92 \pm 0.970	
Ke (h ⁻¹)	Control group	6	0.052 \pm 0.001	0.06
	Second group	6	0.049 \pm 0.012	
$t_{1/2}$ (h)	Control group	6	13.38 \pm 1.82	0.06
	Second group	6	18.97 \pm 5.61	
AUC ₀₋₂₄ (ng/ml.h)	Control group	6	2221 \pm 730	0.59
	Second group	6	1975 \pm 333	
AUC _{0-∞} (ng/ml.h)	Control group	6	3069 \pm 808	0.55
	Second group	6	2747 \pm 497	
MRT (h)	Control group	6	9.22 \pm 0.62	0.55
	Second group	6	8.79 \pm 0.83	

Control group: Rabbits received CsA (7.5mg/kg/day) orally for five days; Second group: Rabbits received CsA (7.5mg/kg/day) concurrently with DEX (0.66mg/kg/day) orally for five days; Statistical significance: $P \leq 0.05$.

In this study, C_{max} of CsA (control group) was 254.9 ± 106.4 ng/mL, and t_{max} was 1.50 ± 0.87 h. The rabbits treated with DEX 0.33 mg/kg/day (first group) produced C_{max} and t_{max} of 290.0 ± 144.9 ng/mL and 1.83 ± 0.75 h, respectively. A slight increase of C_{max} and t_{max} of CsA were found when DEX (first group) was co-administered with CsA compared to the control group. The differences were statistically insignificant. A slight decrease in AUC_{0-24} and $AUC_{0-\infty}$ was manifested in the first test group in comparison to control, but it was statistically insignificant ($P \geq 0.05$). Other PK parameters, including $t_{1/2}$ and K_e were also insignificant (Table 1). Blood concentration-time profiles of CsA were comparable for control, and second test groups (Figure 1).

PK parameters of CsA (C_{max} , t_{max} , k_e , $t_{1/2}$, AUC_{0-24} , $AUC_{0-\infty}$ and MRT) were unaffected ($P \geq 0.05$) by co-administration of DEX at a concentration of 0.66 mg/kg/day (second group) as shown in (Table 2).

3. DISCUSSION

Most immunosuppressive agents have narrow therapeutic indexes, so dosing most of the immunosuppressive agents is applied under careful monitoring of their blood concentrations. Knowing the potential factors that can modify immunosuppressive therapy and pharmacokinetics and metabolic drug interactions can decrease the fluctuation of immunosuppressant blood concentrations¹⁷. CsA is extensively metabolized in the liver by the CYP3A4 system, a member of the CYP450 family of oxidizing enzymes^{3,12}. Sustained and clinically significant DDIs can occur during long-term therapy with CsA. The co-administration of multiple drugs with CsA could result in graft rejection, renal dysfunction, or other undesirable effects. Potential CsA drug interaction is of great clinical importance⁴. Drugs affecting

CYP3A4 metabolic activity are a possible candidate for such interactions¹⁷. Synthetic glucocorticoids as dexamethasone or prednisolone are known substrates and inducers of CYP3A enzymes¹⁸. The regulation of these enzymes, particularly CYP3A4, has been extensively studied¹⁹. In vitro studies of DEX in cultures of human hepatocytes showed potent CYP3A4 inducing effects. In vivo studies had shown that co-administration of DEX with some drugs as phenytoin resulted in reduced plasma level of the drug. When DEX was discontinued, plasma concentration increased by 300% and increased CYP3A4 activity in healthy volunteers and human hepatocyte cultures²⁰⁻²⁵. The conducted study of DEX-CsA interaction showed statistically insignificant differences in PK parameters when CsA was administered alone or in combination with DEX given at two different doses. Similar results were obtained in another research work realized by Villikka and collaborators who studied the PK of triazolam (CYP3A4 substrate) when co-administered with DEX (4-day course of 1.5 mg dexamethasone daily)²⁶. The impact of an enzyme inducer like DEX increases gradually over time. The effect of DEX doses on PK of tacrolimus was manifested after three months of treatment after one month²⁷. This could explain the obtained results since DEX was co-administered with CsA for five days. The possibility that higher doses of dexamethasone could induce CYP3A4 and thereby cause clinically significant drug interactions cannot be excluded. This is particularly relevant in cancer chemotherapy as many anticancer agents are CYP3A4 substrates, and DEX as an antiemetic agent is used at high doses²⁸.

In conclusion, the present study results demonstrated that concurrent use of DEX at the examined regimen with CsA had not influenced PK parameters of CsA. Furthermore, PK studies of CsA using DEX at higher doses and for a longer duration and other clinically relevant CYP inducers or inhibitors are advised.

Conflict of interest

The authors proclaim no conflict of interest.

REFERENCES

1. Laurence L., Brunton. *Goodman and Gilman's the pharmacological basis of therapeutics*. 2006; 11th Ed. McGraw-Hill.
2. Mukherjee S., Mukherjee U.A. comprehensive review of immunosuppression used for liver transplantation, J. Transplant. 2009; 1-20.
3. Zheng S., Tasnif Y., Hebert MF., Davis CL., Shitara Y., Calamia JC., Lin YS., Shen DD., Thummel KE. CYP3A5 gene variation influences cyclosporine A metabolite formation and renal cyclosporine disposition. *Transplantation*. 2013; 95(6): 821–827.
4. Naesens M., Kuypers D., Sarwal M. Calcineurin Inhibitor Nephrotoxicity. *CJASN*. 2009; 4(2): 481-508.
5. Palleria C., Di Paolo A., Giofrè C., Caglioti C., Leuzzi G., Siniscalchi A., De Sarro G., Gallelli L. Pharmacokinetic drug-drug interaction and their implication in clinical management. *J Res Med Sci*. 2013; 8(7): 601–610.
6. Christians U., Sewing K.F. Cyclosporine metabolism in transplant patients. *Pharmacol Ther*. 1993; 57: 291-345.
7. Alloway R.R., Isaacs R., Lake K., Hoyer P., First R., Helderman H., Bunnapradist S., Leichtman A., Bennett M.W., Tejani A., Takemoto S.K. Report of the american society of transplantation conference on immunosuppressive drugs and the use of generic immunosuppressants. *Am. J. Transplant*. 2003; 3: 1211-1215.
8. Fatouros DG., Karpf DM., Nielsen FS., Mullertz A. Clinical studies with oral lipid-based formulations of poorly soluble compounds. *Ther Clin Risk Manag*. 2007;3(4):591-604.
9. Faulds D., Goa K.L., Benfield P. Cyclosporine: a review of its pharmacodynamic and pharmacotherapeutic properties and therapeutic use in immunoregulatory disorders. *Drugs*. 1993; 45: 953-1040.
10. Cook A.M., McDonnell A.M., Lake R.A., Nowak A.K. Dexamethasone co-medication in cancer patients undergoing chemotherapy causes substantial immunomodulatory effects with implications for chemotherapeutic strategies. *Oncoimmunology*. 2016; 5(3): 1-11.
11. Zanger U.M., Schwab M. Cytochrome p450 enzymes in drug metabolism: regulation of gene expression, enzyme activities, and impact of genetic variation. *Pharmacology and Therapeutics*. 2013; 138(1):103-141.
12. Ortiz de Montellano PR. Cytochrome P450-activated prodrugs. *Future Med Chem*. 2013;5(2):213-228.
13. Sahi J.J., Black C.B., Hamilton G.A., Zheng X., Jolley S., Rose K.A., Gilbert D., Le Cluyse E.L., Sinz MW. Comparative effects of thiazolidinedione's on in vitro p450 enzyme induction and inhibition. *Drug Metabolism and Disposition*. 2003; 31: 439–446.
14. Wilkinson G. Drug metabolism and variability among patients in drug response. *The New England Journal of Medicine*. 2005; 352: 2211-2221.
15. Sevrioukova I.F., Poulos T.L. Understanding the mechanism of cytochrome p4503a4: recent advances and remaining problems. *Dalton Trans*. 2013; 42(9): 3116-3126.
16. Sa`adeh A., Aburjai T., ShraimN., Al-Ghazawi M. The Effect of Brassica Oleracea (Cabbage) on The Pharmacokinetics of Ciprofloxacin in an Animal Model. *Jordan Journal of Pharmaceutical Sciences*. 2019; 12(2): 59-67.
17. Parasuraman S., Raveendran R., Kesavan R. Blood sample collection in small laboratory animals, *Journal of Pharmacology and Pharmacotherapeutics*. 2010; 1(2): 87–93.
18. Namburu K., Kolapalli V., Prasanna Raju Yalavarthi R., Harini Chowdary Vadlamudi H., Jaya Preethi Peesa J. Pharmacokinetic Profiling and Bioavailability Assessment of Meloxicam Solid Dispersion Tablets. *Jordan Journal of Pharmaceutical Sciences*. 2018; 11(2): 85-92.
19. Monostory K. Metabolic Drug Interactions with Immunosuppressants. In: Tsoulfas G. *Organ Donation and Transplantation-Current Status and Future*

- Challenges*. 2018; 409-440.
20. Lam S., Partovi N., Ting LS., Ensom MH. Corticosteroid interactions with cyclosporine, tacrolimus, mycophenolate and sirolimus: fact or fiction? *Ann Pharmacother*. 2008; 42(7):1037-1047.
 21. Guengerich F.P. Cytochrome P-450 3A4: regulation and role in drug metabolism. *Annu Rev Pharmacol Toxicol*. 1999; 39: 1-17.
 22. Coutinho A.E., Chapman K.E. The anti-inflammatory and immunosuppressive effects of glucocorticoids, recent developments and mechanistic insights. *Mol. Cell Endocrinol*. 2011; 335(1): 2-13.
 23. Pascussi J.M., Drocourt L., Gerbal-Chaloin S., Fabre J.M., Maurel P., Vilarem M.J. Dual effect of dexamethasone on CYP 3A4 in human hepatocytes. Sequential role of Glucocorticoid receptor and Pregnane X receptor. *Eur J Biochem*. 2001; 268(24): 6346-6358.
 24. McCune J.S., Hawke R.L., LeCluyse E.L., Gillenwater H.H., Hamilton G., Ritchie J., Lindley C. In vivo and in vitro induction of human cytochrome CYP450 3A4 by dexamethasone. *Clin Pharmacol Ther*. 2000; 68: 356-366.
 25. Lin J.H., Chiba M., Chen I.W., Nishime J.A., deLuna F.A., Yamazaki M., Lin Y.J. Effect of dexamethasone on the intestinal first-pass metabolism of indinavir in rats: Evidence of cytochrome CYP 4503A and P-Glycoprotein induction. *Drug Metab Dispos*. 1999 ; 27(10): 1187-1193.
 26. Chiba M., Hensleigh M., Lin J.H. Hepatic metabolism of indinavir, an HIV protease inhibitor in rat and in human microsomes. Major role of CYP3A. *Biochem Pharmacol*. 1997; 25: 53(8): 1187-1195.
 27. Lackner T.E. Interaction of dexamethasone with phenytoin. *Pharmacotherapy*. 1991; 11(4): 344-347.
 28. Villikka K., Kivistö K.T., Neuvonen P.J. The effect of dexamethasone on the pharmacokinetics of triazolam. *Pharmacol Toxicol*. 1998; 83: 135-138.
 29. Anglicheau D., Flamant M., Schlageter M., Martinez F., Cassinat B., Beaune P., Legendre C., Thervet E. Pharmacokinetic interaction between corticosteroids and tacrolimus after renal transplantation. *Nephrology Dialysis Transplantation*. 2003; 18(11): 2409-2414.
 30. Grunberg S.M. Antiemetic activity of corticosteroids in patients receiving cancer chemotherapy: dosing, efficacy, and tolerability analysis. *Annals of Oncology*. 2007; 18: 233-240.

تأثير دواء الديكساميثازون بجرعات مختلفة على حركية دواء السيكلوسبورين

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ملخص

تم تصميم هذه الدراسة للتحقيق في وجود تفاعل حركي دوائي بين ديكساميثازون (DEX) بتركيزات مختلفة على حركية دواء السيكلوسبورين (CSA) في الأرانب الذكور السليمة صحياً. تم اختيار الأرانب وتقسيمها إلى ثلاث مجموعات: المجموعة الضابطة (ع = 6) تلقت محلول السيكلوسبورين عن طريق الفم (7.5 ملغم / كجم / يوم) بعد تحضيره من كبسولة الجيلاتين الطرية لمدة خمسة أيام وفي اليوم الخامس تم سحب عينات الدم التسلسلية من الوريد الطرفي لاذن الأرانب في فترات زمنية مختلفة بعد الجرعات. أعطيت الأرانب في المجموعتين الأولى والثانية عن طريق الفم محلول السيكلوسبورين (7.5 مجم / كجم / يوم) وديكساميثازون بالتزامن مع جرعتين (0.33 و 0.66 مجم / كجم / يوم) على التوالي. في اليوم الخامس من اعطاء الدوائين ، تم جمع عينات دم متسلسلة لكل مجموعة اختبار على مدى 24 ساعة كما في المجموعة الضابطة ومن ثم تم تحديد الحركيات الدوائية للسيكلوسبورين في مجموعات الضبط والاختبار. لوحظ أنه كانت هناك اختلافات غير مهمة احصائياً بين مجموعات الضبط والاختبار عند اعطاء محلول السيكلوسبورين مع الديكساميثازون بالتركيزين المذكورين اعلاه مما يشير إلى أن الديكساميثازون لا يؤثر على حركية دواء السيكلوسبورين.

الكلمات الدالة: السيكلوسبورين. الديكساميثازون. التفاعلات الدوائية . المعلمات الحركية الدوائية.

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